

Using NMR to study Macromolecular Interactions

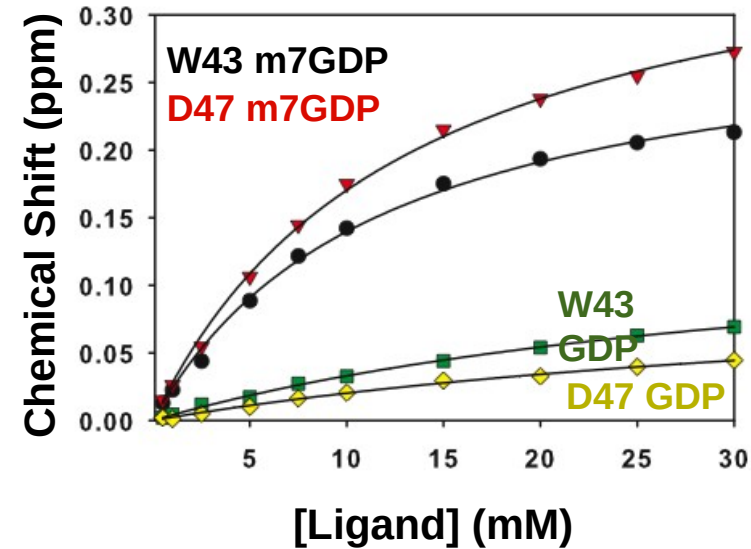
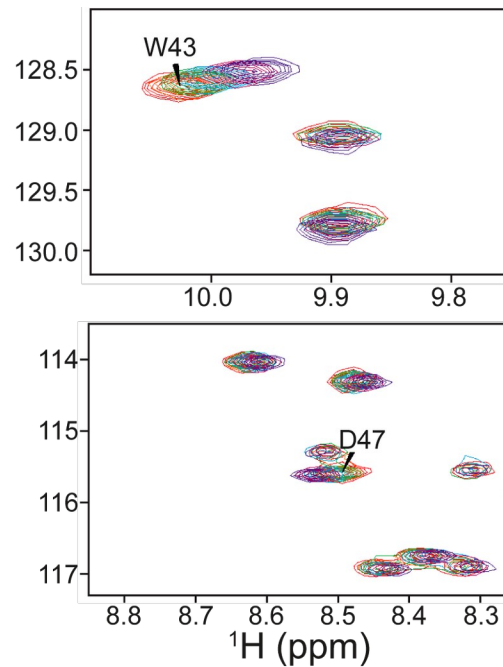
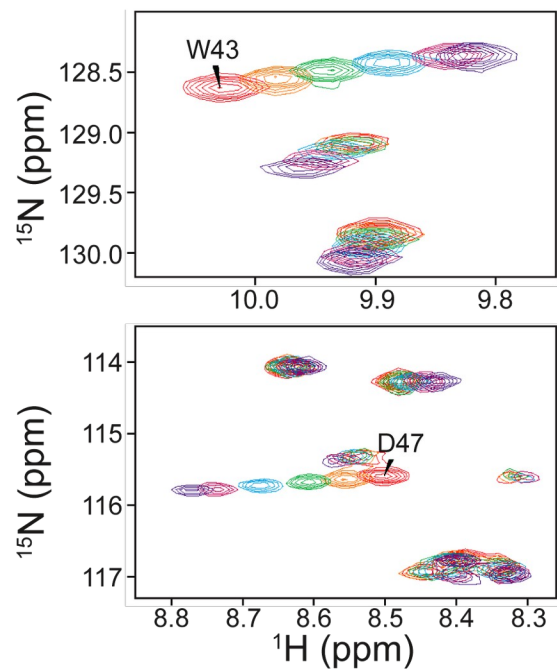
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Outline

- Multidimensional NMR
- Macromolecular Interactions
- Dynamics
- Dealing with large complexes
- Structure Determination

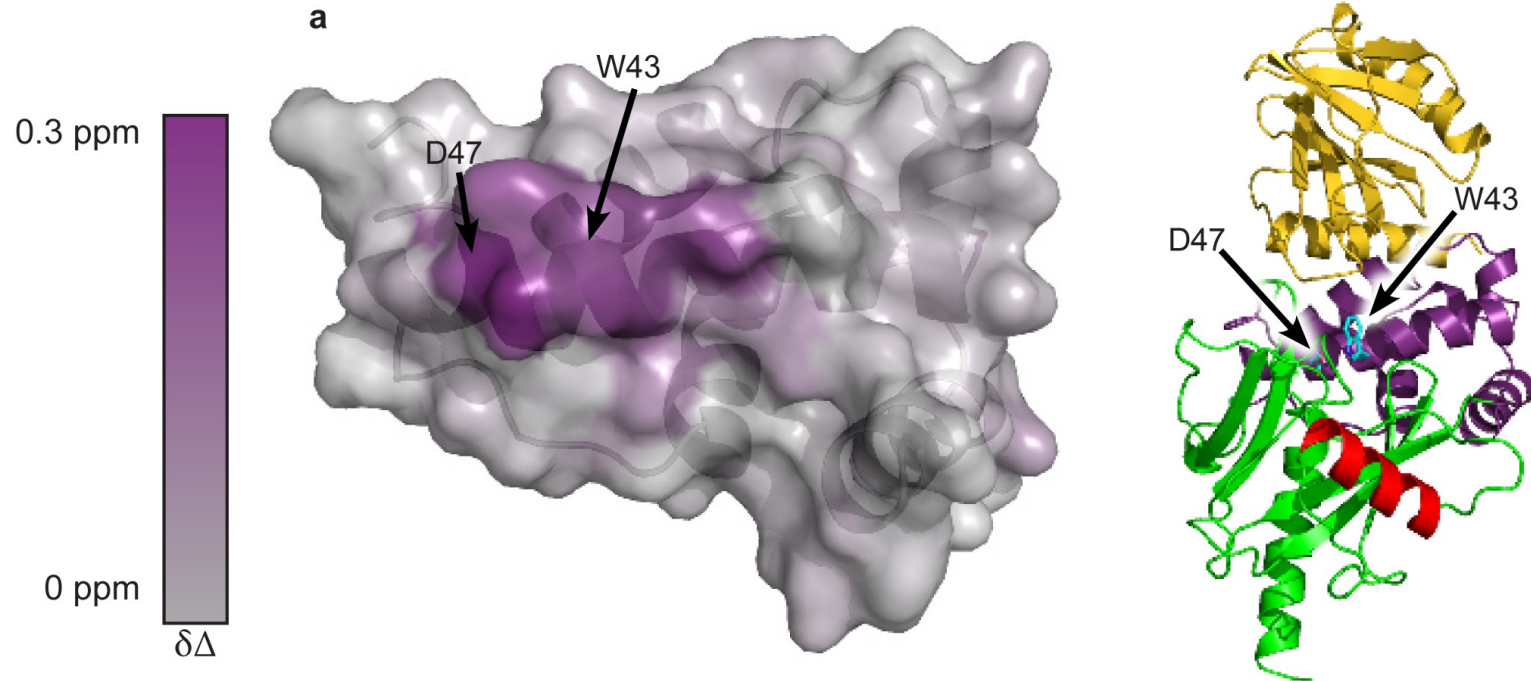
Part II:
Macromolecular Interactions Detected
by NMR

Binding of nucleotide to protein



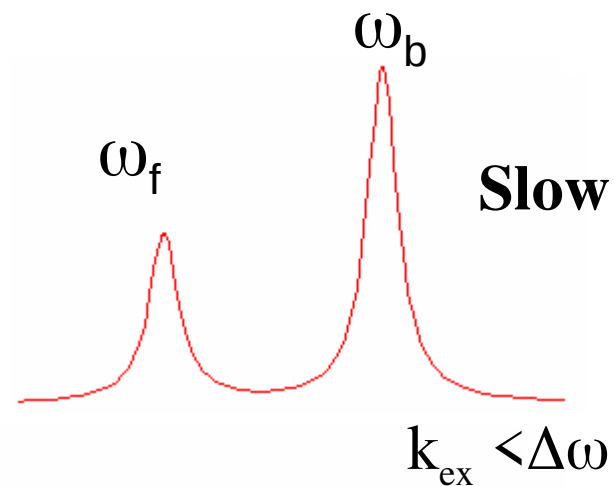
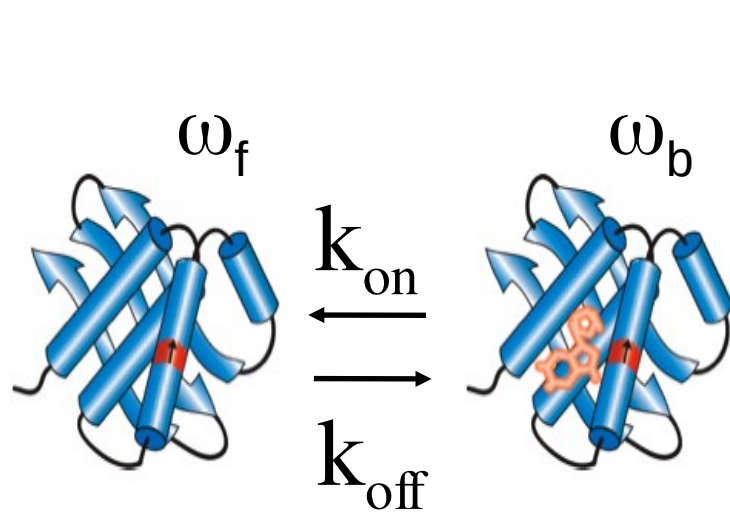
Dose dependent resonance shifts can be fit to obtain K_d

Shifts may be color coded onto surface to identify ligand binding site



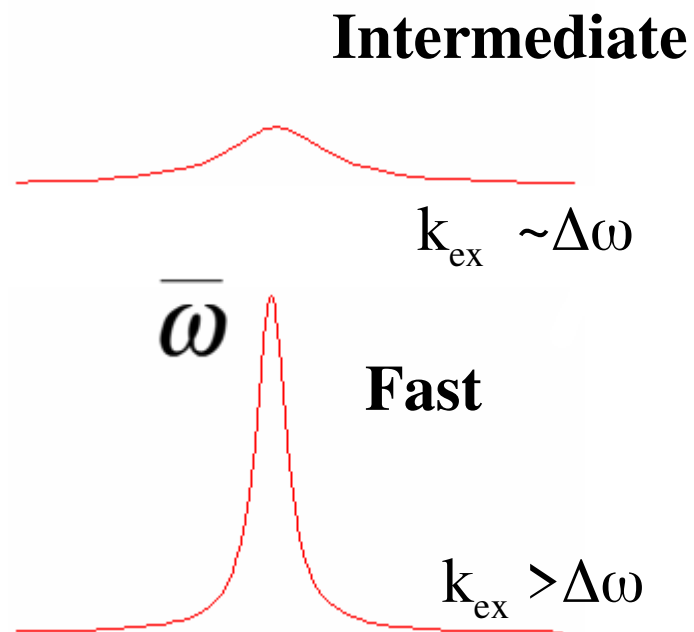
Caveats?

NMR to monitor ligand binding



$$k_{ex} = k_{on}[L] + k_{off}$$

$$\Delta\omega = \omega_f - \omega_b$$

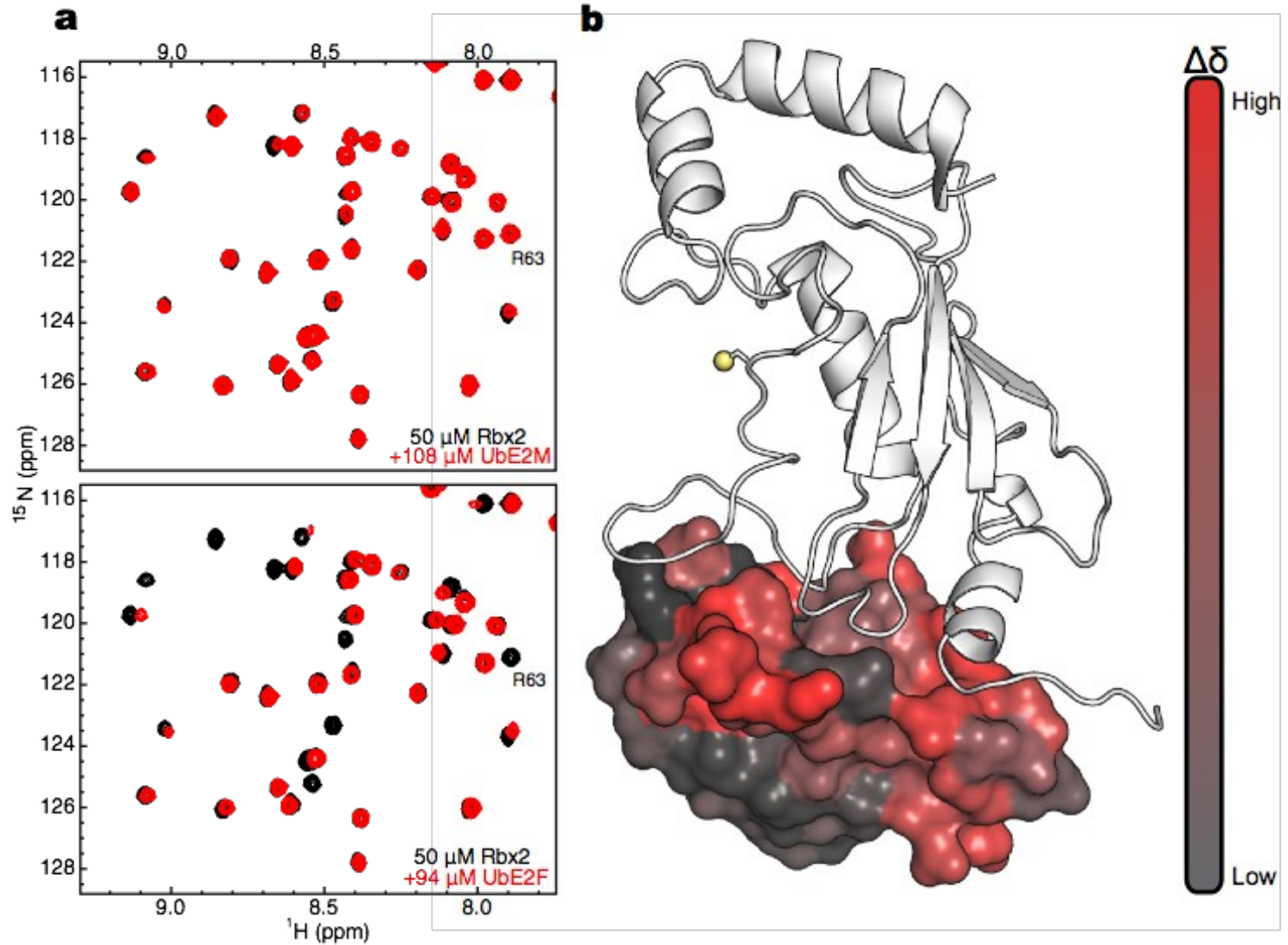


Fraction bound of labeled protein

$$P_b = \frac{\bar{\omega} - \omega_f}{\omega_b - \omega_f} = \frac{[L]}{[L] + K_d}$$

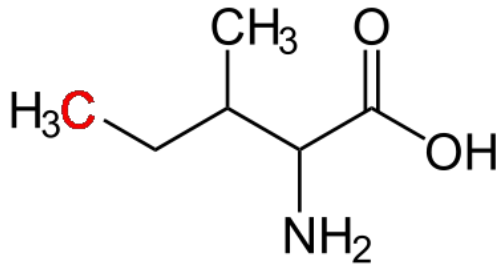
$\bar{\omega}$: observed chemical shift

Monitoring Protein/Protein Interactions by HSQC

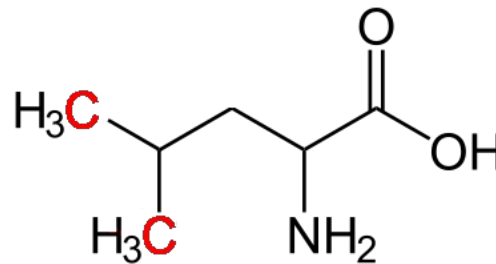


Sparse Labeling to Simplify Spectra

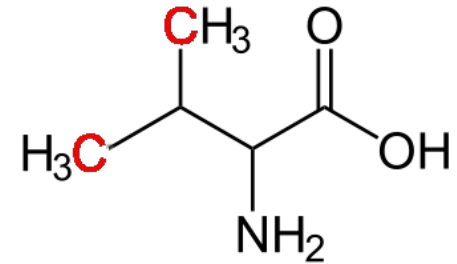
Selectively label R group methyls with C-13 (NMR visible)



Isoleucine



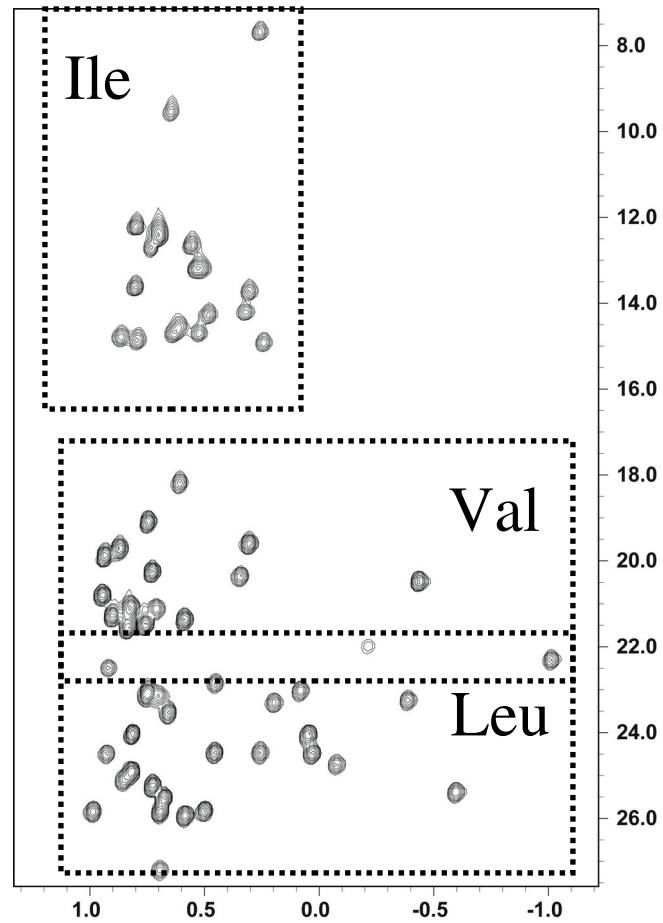
Leucine



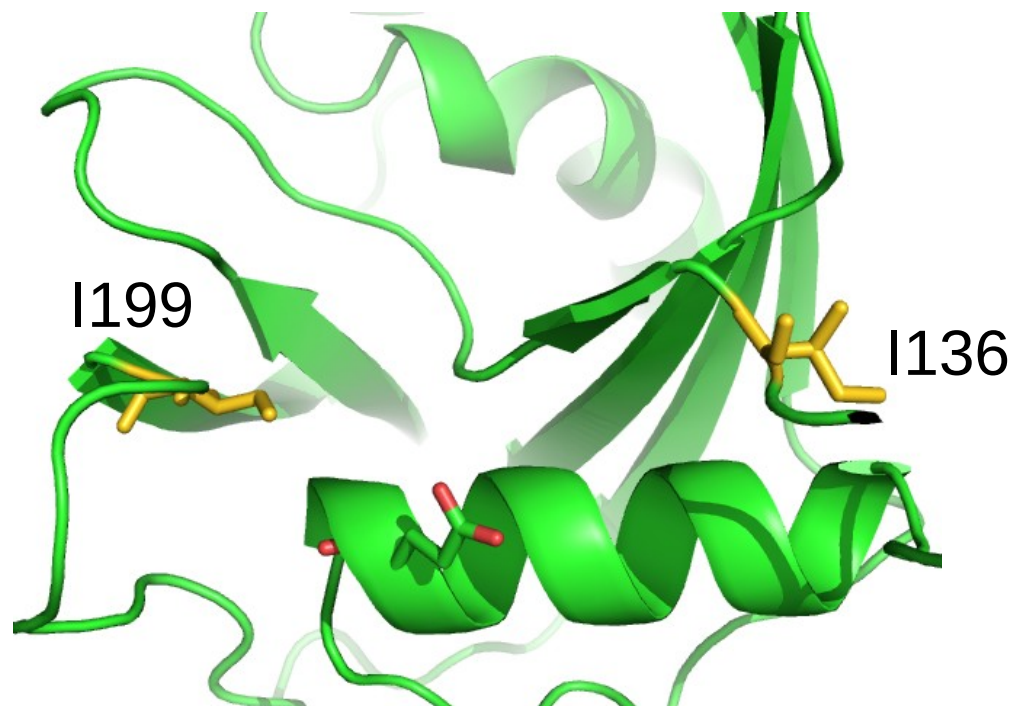
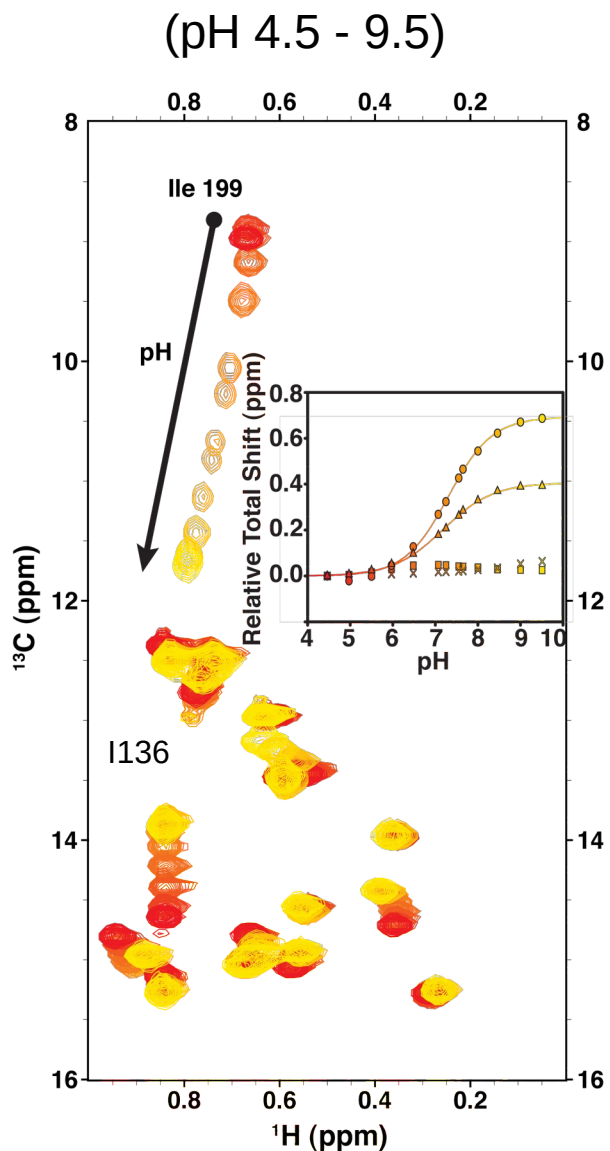
Valine

(add alpha-ketoacid precursors to ILV 30 minutes prior to induction)

^{13}C HSQC of ILV labeled protein

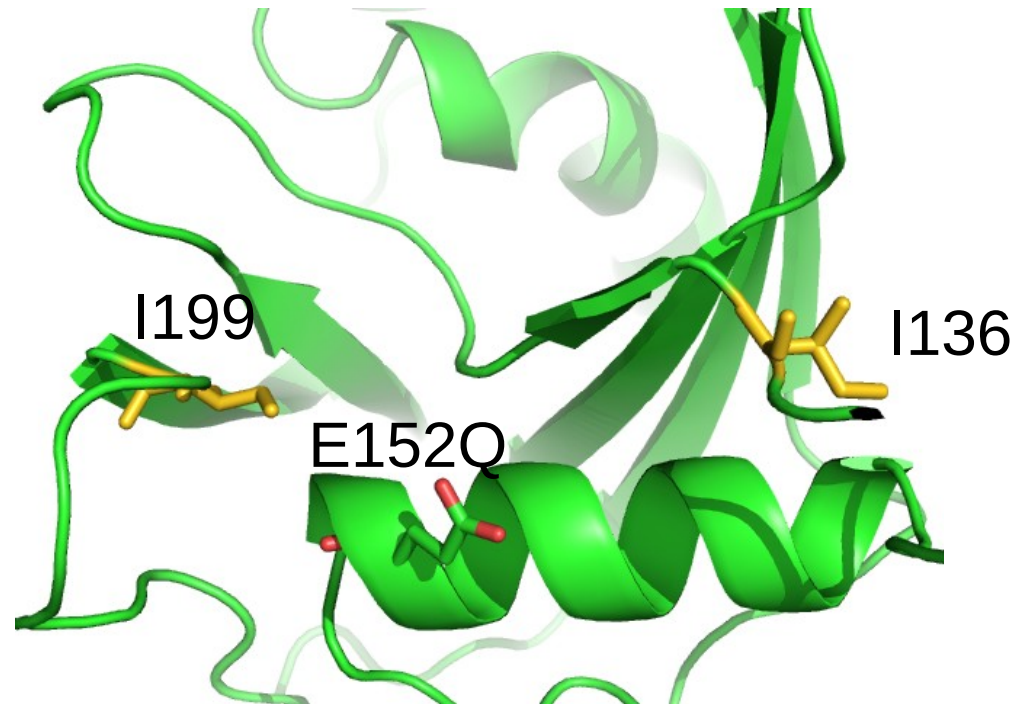
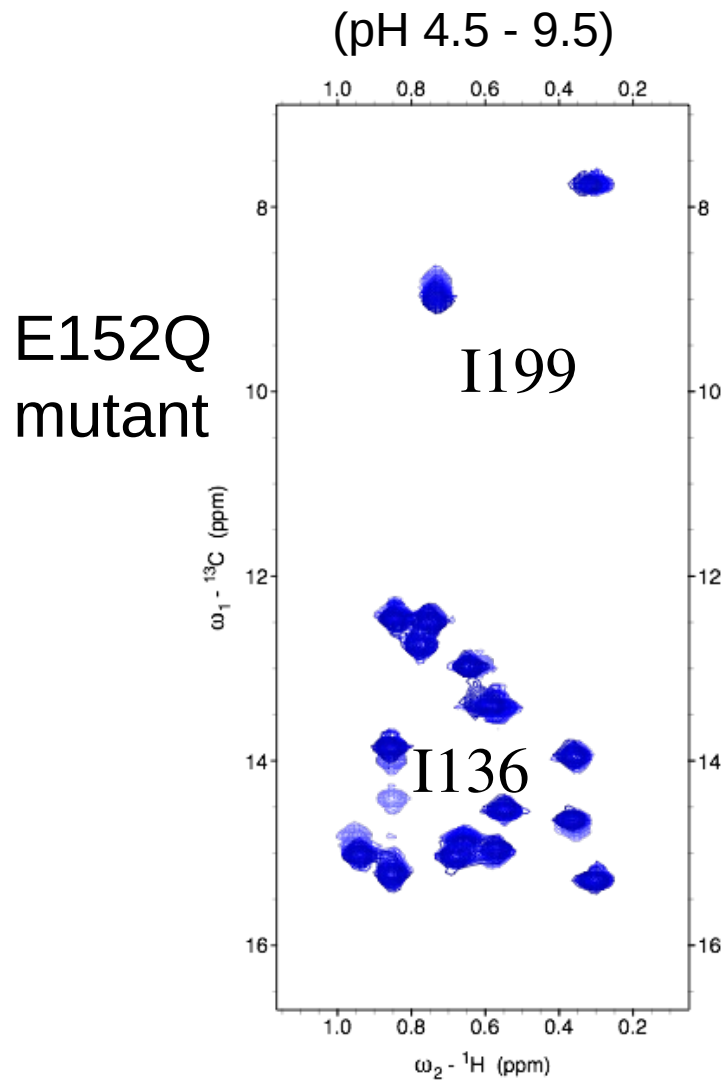


Measuring pK_a by NMR

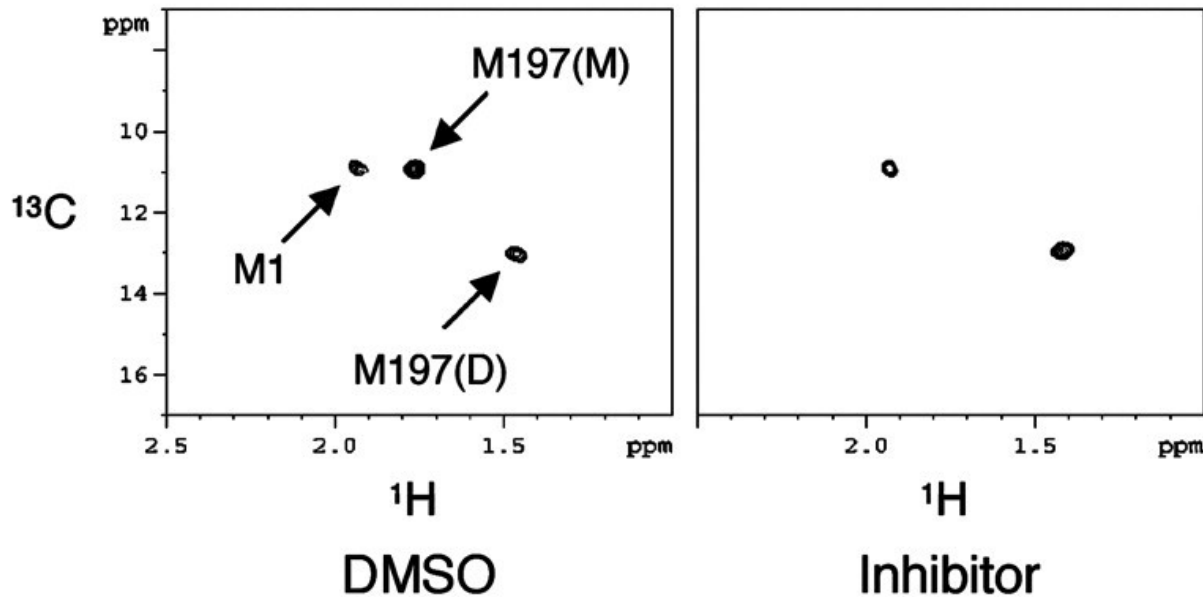


pK_a of 7.2, elevated for Glu

*Identification of titratable residue by
site-directed mutagenesis and NMR*



*Example of slow exchange:
monomer-dimer equilibrium*



**Inhibition of KSHV Pr
stabilizes
the dimeric conformation**

Methionine specific labeling simplifies analysis