Bi 204 Methods: X-Ray diffraction . Calibrating Non-covalent Molecular Interactions

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A 'Ligand' the cancer drug imatinib (Gleevec) bound to the tyrosine kinase Abl.

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Optical image formation, - without lenses









If automated- why are there errors? What do I trust? Examples of errors trace sequence backwards, mis assignment of helices etc

Automated Crystal Growth



Structure Solution



Database

NH3 sites and the role of D160 at 1.35Å Resolution



Data/Parameter ratio is the same for all molecular sizes at the same resolution d_{min} ie. quality is the same!





Determining Atomic Structure

- X-ray crystallography = optics $\lambda \sim 1.5$ Å (no lenses)
- Bond lengths ~1.4Å
- Electrons scatter X-rays; X-rays 'see electrons'
- Resolution –Best is $\lambda/2$ Typical is 1 to 3 Å
- Accuracy of atom center positions ±1/10 Resolution



CCD detector



Resolution $d_{min} = \lambda / 2 \sin(\theta_{max})$ differs from Rayleigh criterion



The Rayleigh Criterion

• The Rayleigh criterion is the generally accepted criterion for the minimum resolvable detail - the imaging process is said to be diffraction-limited when the first diffraction minimum of the image of one source point coincides with the maximum of another.





How do we judge the Quality of structure?

2. Overall quality criteria: agreement of observations with diffraction calculated from the interpreted structure.

3. Since we refine the structure To match the I_{hkl} overfitting ?

Define R_{free} for a 'hold-out ' set of observations.

¹/₂ 4. OK? R < 20%, R free< 25%

5. But the experimental errors in measuring Fo are ~ 3%. inadequate models of solvent, atom motion, anharmonicisity

6 Accuracy ~ 0.5*res*R

Scattering by matter - (interference) of a single wavelength Xray Scattering from a point is equal in all directions. add a second point, scattering in some direction <u>si</u> The second wave, seathered by B travels further by the distance PB + BQ. Ik scattered wave lags in phase by = $\frac{2\pi}{\lambda}$ ([P-B] + [B-Q]) where \$= path length estra for B versus the reference A. = 2 () ©Robert M. Stroud 2017

Adding up the scattering of Atoms: 'interference' of waves



Constructive Interference Destructive Interference

Waves add out of phase by $2\pi[\text{extra path}/\lambda]$



In general they add up to something amplitude In between -2f and +2f. For n atoms

 $F^2 = f \times \sqrt{n}$



of amplitude less than 2, (ie < the number of points) $A_{mp} = |F| = 2 \cos 2\pi \left(\frac{\Phi}{2\lambda}\right)$ which has the same periodicity in time $\sin 2\pi \left(\frac{x+\overline{b}/2}{\lambda} - \omega t\right)$ and is 'phase shifted' by \$1/2

We only observe intensity of scattering = (Auplitude), - or the number of photons scattered of any one position. Thus the "time" dependent consoment is irrelevant. But, the relative "phase shift" is crucial. If the incoming wave is represented by a line = amplitude along the x - axis, -The scattered wave can be represented by the sum of another arrow that represents the scattered wave from A, plus another from B. If X-rays, the amplitude of each component is the normber of electrons in that atom, f:



Just 2 atoms...



to the 1st atom. . ₹/λ Many atoms add by the same rules. Different in every direction. E has the required amp. & phase. If we use this method we can add i=1 to n different atoms; each amplitude fi Diffracted X-ray beam −fs, Crystallized molecule each atom has fi cos x; along z and fi sin x; alongy Film If we put 'mits' on the axes, we can add up the 'sc' and 'y' components to write the sum over "x"; the sum over "y", -hence colculate F as a wave of amplitude $|\mathbf{F}| = \sqrt{\left(\sum_{i \in f_i} \cos x_i\right)^2 + \left(\mathbf{E}_i \mathbf{f}_i \sin x_i\right)^2}$ and $\alpha = \tan^{-1}\left(\frac{\sum f_i \sin \alpha_i}{\sum f_i \cos \alpha_i}\right)$ ©Robert M. Stroud 2017 13



Data/Parameter is the same for all molecular sizes at the same resolution d_{min}

ie. quality is the same!

1/Resolution



duce the lattice of 12. Subsidiary diffraction maxima due to the small humber of apertures can be seen clearly.

Plate 10



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Ø





This is all there is? YES!!

FT⁻¹

FT

Scattering pattern is the Fourier transform (FT) of the structures Amplitude and phase of waves is a sum of waves from each atom

 $\mathbf{F}(\underline{\mathbf{S}}) = \Sigma_j \mathbf{f}_j \mathbf{e}^{(2\pi i \mathbf{r}_j \cdot \mathbf{S})}$

Structure is the 'inverse' Fourier transform of the Scattering pattern





Scattering pattern is the Fourier transform of the structure

 $\mathbf{\underline{F}(\underline{S})} = \Sigma_j \mathbf{f}_j \mathbf{e}^{(2\pi i \mathbf{r} \mathbf{j}. \mathbf{S})}$

Structure is the 'inverse' Fourier transform of the Scattering pattern



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The sequence shows how the diffraction pattern of a simple object is built up by superposition of sets of fringes.

 $f(\underline{c})$ Plate 2 Two atoms One atom



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Plate 11	Object	Build a d	crystal	Scattering	Plate 11	
· · ••	8.0	° °	1 1 2 000 p 10 00 p 10 0 p	₩. 4₩-2₩ 1		
ê ê	ш ф р ф	8 ° ¤ °			2011年 1999年 1999年 1999年 1999年 1999年 1997 1997	
	$ \begin{array}{cccccccccccccccccccccccccccccccccccc$					

$$\underline{\mathbf{F}}_{(\mathbf{h},\mathbf{k},\mathbf{l})} = \sum_{j} \mathbf{f}_{j} \mathbf{e}^{(2\pi \mathbf{i} (\mathbf{h} \mathbf{x}_{j} + \mathbf{k} \mathbf{y}_{j} + \mathbf{l} \mathbf{z}_{j}))}$$

Every X-ray reflection (h,k,l) has a contributing wave from all j atoms .

$$\rho(\mathbf{x},\mathbf{y},\mathbf{z}) = \Sigma_{hkl} \quad \underline{\mathbf{F}}_{(\underline{h},\underline{k},\underline{l})} e^{(-2\pi i(hx+ky+lz))}$$

or
$$\rho(\mathbf{x},\mathbf{y},\mathbf{z}) = \Sigma_{hkl} \mid \underline{\mathbf{F}}_{(\underline{h},\underline{k},\underline{l})} \mid e^{(-2\pi i(hx+ky+lz) + \phi_{hkl})}$$

Every point in the density map has contributions from every reflection

PROOF OF INVERSE FOURIER TRANSFORM

Once we measure Intensity = |Fe)² we calculate |Fe/ X(s) (it's phase) We need to determine We can invert the "observations", to calculate the 'scattering density' map. For X-rays = electron density map. by $\rho(\mathbf{\underline{r}}) = \sum \underline{F}(\underline{\mathbf{s}}) \ e^{-2\pi i \, \mathbf{\underline{r}}_i \cdot \underline{\mathbf{s}}} = \int \underline{F}(\underline{\mathbf{s}}) \ e^{-2\pi i \, \mathbf{\underline{r}}_i \cdot \underline{\mathbf{s}}}$ Just as all atoms contribute to each amplitude F(s) so each F(s) contributes to p(c). called "Fourier inversion". Note rectors are nnderlined Proof? postulate true, then apply for some s' F(s) =) () F(s) e-zui [: sd) e zui r's' dVr $= \int_{S} \int_{S} F(s) e^{2\pi i s' - s} ds dt$ This s = 0 unless s' = s, then = F(s) = ©Robert M. Stroud 2017 QED

Scattering pattern is the Fourier transform of the structure

 $\mathbf{F}(\underline{\mathbf{S}}) = \Sigma_j \mathbf{f}_j \mathbf{e}^{(2\pi i \mathbf{r} \mathbf{j}. \mathbf{S})}$

Structure is the 'inverse' Fourier transform of the Scattering pattern

 $\rho(\underline{\mathbf{r}}) = \sum \mathbf{F}(\underline{\mathbf{S}}) e^{(-2\pi i \mathbf{r}.\mathbf{S})}$

- 'Difference maps'
- -Define bound ligands
- -to find any missing atoms during refinement,
- -to find ligands
- -define movements of protein or water
- -determine ion positions
- -determine changes in dynamic motion

Suppose we interpret 7 atoms; but 3 remain to be found in density



In reality, maybe 3 atoms are missing. How to see what is missing?



and why do we care?

34 a laser beam put waves in phase. $T = (na)^2$ so if a 60 woth lamp bulb produces n photons/sec/Hangle, $I \sim (\sqrt{n} a)^2 = n a^2$. Intensity laser = How much change will a in a 2000 atom profem lys -> met mutation. give? Rg 3 Protein 76 each step amp = 12000 * 7E = 313. (313.05 Sulfur - Carbon = 10 E. ie 3% of the whole protein !! $\frac{But}{(1 + 1)^{2}} = (amp^{2}) = 313.55$ $< Intensity = 2000 \times 7^{2} + 10^{2}$ $= (313.24)^{2}$ But $\Delta amp = 10 \frac{1}{\sqrt{2}} = 7.1 \text{ c}$ AI/T = 4.4 %.

How much difference will it make to the average intensity? average amplitude?

if we add a single Hg atom?

and why do we care?

How much difference will 10 electrons make to the average intensity? 98,000 e² average amplitude? 313

average difference in amplitude? average difference in intensity?

if we add a single Hg atom?

$$\frac{69^2}{2} = a |aser beam put waves in phase.$$

$$I = (n a)^2$$

$$I = (n a)^2$$

$$So \cdot if a 60 wolt lamp bulb
produces n photous /sec/trangle,
$$I \sim (\sqrt{n} a)^2 = n a^2.$$

$$Intensity |aser = n$$

$$I |aser = n$$

$$\frac{1}{2} |aser = n$$

$$\frac{$$$$

and why do we care?

or Hg atom n=80e

How much difference will 80 electrons make to the average intensity? 98,000 e² average amplitude? 313 e

average difference in amplitude?18 % of each amplitude!36% of each Intensity

difference in 'intensity?' 104,400-98000=6400 (6.5%)

$$\frac{22}{3} = a \operatorname{laser} \operatorname{pert} \operatorname{waves} \operatorname{in} \operatorname{phase}.$$

$$I = (n a)^{2}$$

$$I =$$

USES: 2. Add a substrate, Grow a new crystal Measure New $|F(\underline{S})|_{obs+substrate}$ Compare with the apo-protein. Transform $\Delta F = ||F(\underline{S})|_{obs+substrate} - |F(\underline{S})|_{obs} ||\Phi(\underline{S})|_{obs}$ or $[2|\mathbf{F}(\underline{S})|_{obs+substrate} - |\mathbf{F}(\underline{S})|_{obs}] \Phi(\underline{S})$ |**F**(<u>**S**</u>)|_{observed} $= a' 2F_0 - F_0 map'$ t_{true} It is **unbiased** as to where the missing **F**(**s**) substrate is. phase $\Phi(S)$ ©Robert M. Stroud 2017 28

A Dfference map shows 1/3 occupied NH3 sites and the role of D160 at 1.35Å Resolution. Here are 0.3 NH_3 peaks!



Khademi..Stoud 2003

Fo-Fc maps identify everything ordered that is 'missing'

10772 Biochemistry, Vol. 41, No. 35, 2002



Valiyaveetil et al.

10774 Biochemistry, Vol. 41, No. 35, 2002



FIGURE 1: Lipid molecules in KcsA crystals. A stereoview of the KcsA structure with electron density corresponding to the lipid molecule. The backbone of KcsA is shown as a red and yellow trace. Green spheres represent potassium ion binding sites. The $F_o - F_c$ map (contoured at 3σ) was calculated using a model that does not contain lipid molecules. For clarity, density corresponding to only one of the lipid molecules is shown. The KcsA monomer consists of an N-terminal outer helix, a central pore helix, and a C-terminal inner helix. This figure was prepared with MOLSCRIPT (31) and Raster3D (32).

-Eliminate Bias-Half electron content-See electrons

FIGURE 3: Structural analysis of lipid binding to KcsA. (a) Binding surface of the lipid molecule. The surface of KcsA is colored according to curvature (green, convex; gray, concave). The lipid molecule, built as 1,2-diacylglycerol, is shown in CPK representation with oxygen atoms colored red and carbon atoms colored yellow. (b) Lipid-binding site viewed from the extracellular side along the 4-fold axis of KcsA. The channel is colored blue. The green sphere represents the potassium ion. The lipid molecule is in CPK representation colored as in panel a. Panel a was prepared with GRASP (33). Panel b was prepared with MOLSCRIPT (31) and Raster3D (32). The closer you get –the lower the noise. Can see single electrons.



Figure 3 The catalytic triad. **(A) Stereoview displaying Model H superimposed on the 2Fo Fc (**model H phases) at 1 (aqua) and 4 (gold). The densities for C and N in His 64 are weaker than in Asp 32. The Asp 32 CO2 bond at 4 is continuous, while the density for the C and O1 are resolved. (B) Schematic of the catalytic residues and hydrogen bonded neighbors with thermal ellipsoid representation countered at 50% probability (*29*). Catalytic triad residues Ser 221 and His 64 show larger thermal motion than the Asp 32. Solvent O1059 appears to be a relatively rigid and integral part of the enzyme structure. (C) Catalytic hydrogen bond (CHB). A Fo Fc (model H phases) difference map contoured at +2.5 (yellow) and 2.5 (red) and a 2Fo Fc (model H phases) electron density map contoured at 4 (gold). The position of the short hydrogen atom (labeled HCHB) in the CHB is positioned in the positive electron density present between His 64 N1 and Asp 32 O2.



Fig. 7. The peaks associated with His57 on the difference map. The lower peak is negative density (-) while the other one is positive (+). The latter peak is a composite with a solvent molecule density (see text).



FIG. 8. Electron density for His57 in the DIP-trypsin Fourier map, computed for the plane parallel to the imidazole ring.

F10. 7. The peaks associated with His57 on the difference map. The lower peak is negative density (-) while the other one is positive (+). The latter peak is a composite with a solvent molecule density (see text).

TABLE 1

Analysis of Fourier maps

Мар	$\langle F_{obs} angle \ (e)$	σ (e)	Calculated [†] $\langle \Delta \rho^2 \rangle^*$ (e Å ⁻³)	Observed‡ r.m.s. error (e Å ⁻³)	Obser highest (e Å ⁻³)	rved noise 8.D.§	Obser highest (e Å ⁻³)	ved peak s.p.	
BA-trypsin – DIP-trypsin DIP-trypsin	84·7 573·0	2·3 21·0	0-069 0-38	0-059	0.17	2.2	0.75	11	

$$\begin{split} \dagger \ \Delta F : \langle \Delta \rho^2 \rangle &= \frac{1}{2V^2} \sum_{\text{Add}} \Delta F^2 \ (2-m^2), \\ F_{\text{DIPT}} : \langle \Delta \rho^2 \rangle &= \frac{1}{V^2} \sum_{\text{Add}} F^2_{\text{DIPT}} \ (1-m^2), \end{split}$$

(after Henderson & Moffat, 1971).

‡ The observed root mean-square density error is based on a relatively featureless region of the map.

§ s.D., the electron density given as a @Rhinder With real culded r.m.s. error.

Fo-Fc density at 2.5 and 2.0 σ Green = IntB analog complex, Cyan =dUMP-CB3717 complex



Relative Information in Intensities versus phases



Figure 6.1 Relative amounts of information contained in reflection intensities and phases. (*a*) and (*b*) Duck and cat, along with their Fourier transforms. (*c*) Intensity (shading) of the duck transform, combined with the phases (colors) of the cat transform. (*d*) Back-transform of (*c*) produces recognizable image of cat, but not duck. Phases contain more information than intensities. Figure generously provided by Dr. Kevin Cowtan.

Relative Information in Intensities versus phases



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= incorrect structure

= incorrect structure

-incorrect assumption = incorrect structure

-Incorrect phases

-incorrect model

Electron Diffraction

- -Electrons are scattered more than X-rays:
 - Every diffracted beam is a new incident ray!!
- -from Electric fields in atoms
 - Scatter goes up as square root of atom number
- Absorption is high, so very thin crystals only!
 - 100Å -5000Å
- Phasing? Molecular replacement? Guess?
 - because Heavy metals are less different.
- Tilting specimen to 'scan' diffraction

Images of lysozyme microrystals.(A) Light micrograph showing lysozyme microcrystals (three examples indicated by arrows) in comparison with larger crystals of the size normally used for X-ray crystallography.



Dan Shi et al. eLife Sciences 2013;2:e01345



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MicroED structure of lysozyme at 2.9 Å resolution.(A) The 2Fobs-Fcalc (contoured at 1.5σ) map covers protein residues 5–45 of lysozyme.



Dan Shi et al. eLife Sciences 2013;2:e01345



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Supplementary Proof of the 'Random Walk' calculation

The 'Random Walk' problem? (p33.1-33.3)

What is the average sum of n steps in random directions?

(What is the average amplitude <|F(s)|> from an n atom structure?)

-AND why do we care?!.....

How much difference from adding a mercury atom (f=80).

The average intensity for an n atom structure, each of f electrons is <l>= nf²

The average amplitude is Square root of n, times f

50

$$\left\{ OB \right\}^{2} = \left(\sum_{i=1}^{n} e^{2\pi i q_{i}} \right) \left(\sum_{j=1}^{n} e^{2\pi i q_{j}} \right) \cdot \int^{2} \int^{2} e^{2\pi i (q_{i} - q_{j})} \cdot \int^{2} \int^{2} \int^{2} \int^{2} e^{2\pi i (q_{i} - q_{j})} \cdot \int^{2} \int$$