

# Assignments for Next Monday:

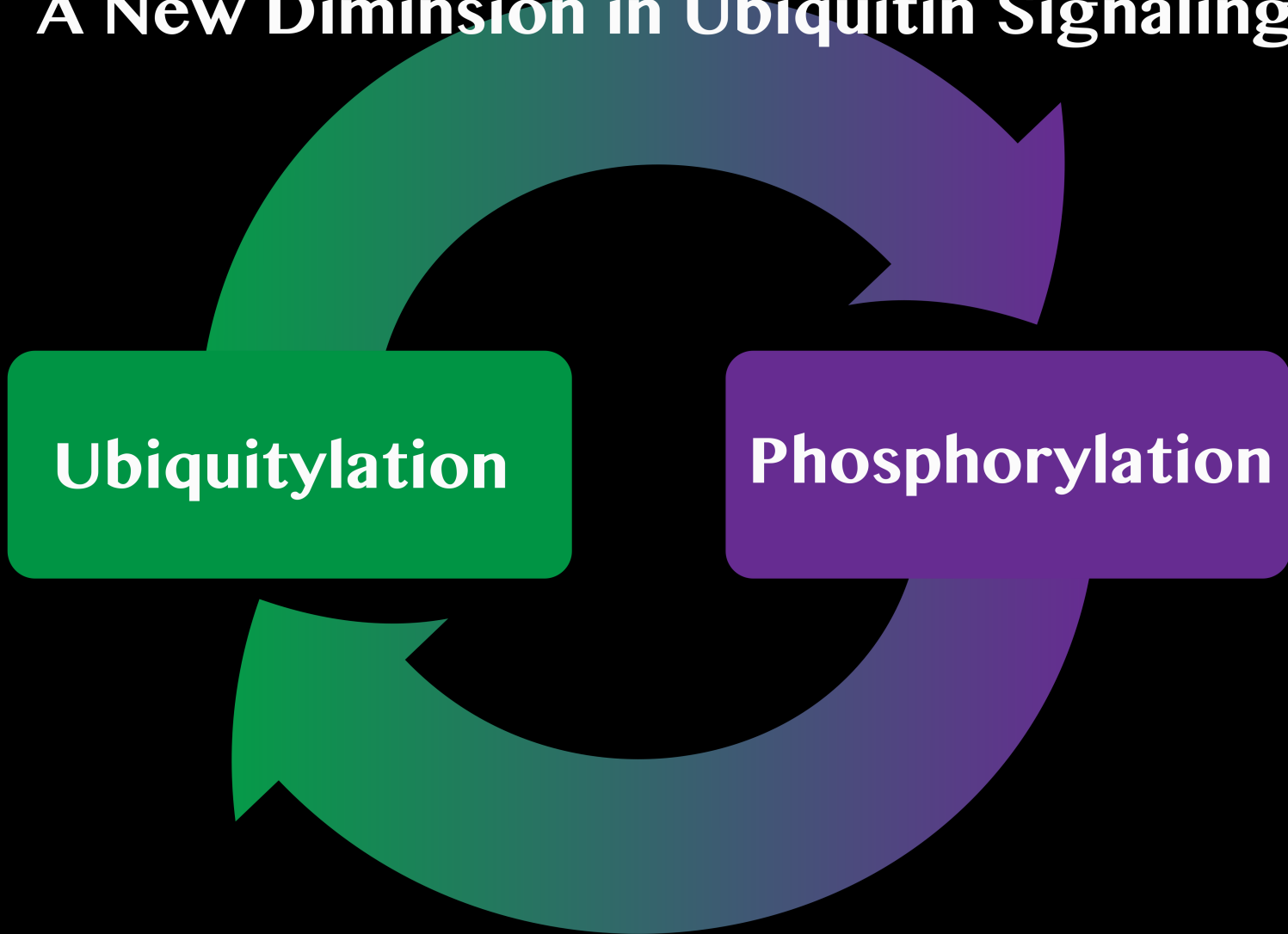


Nathan (Lab work Day 1, 9/28)



Ryan (Journal Club on [Wauer et al, 2015], 9/28)

# Phosphorylation of Ubiquitin: A New Diminsion in Ubiquitin Signaling

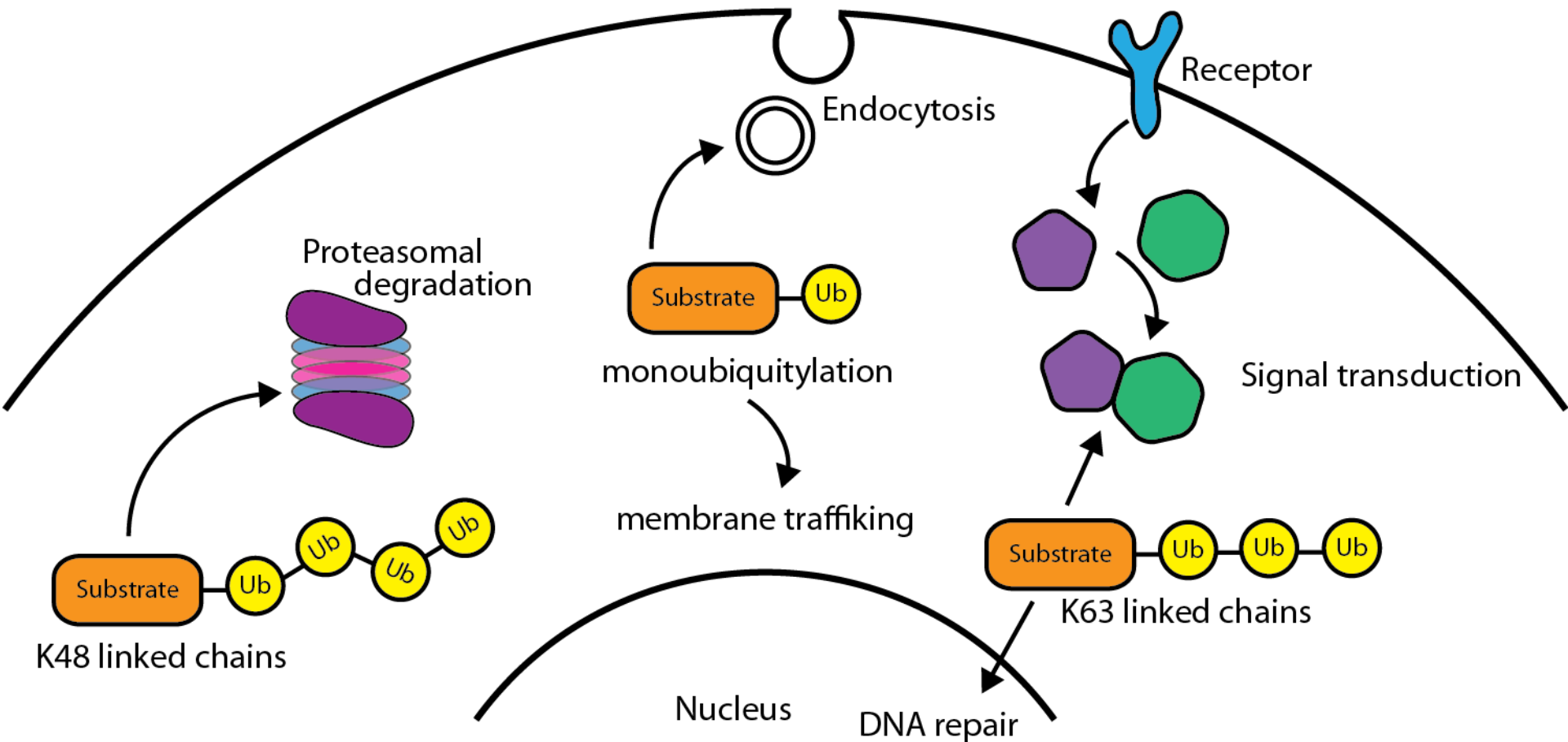


**Danielle Swaney**  
Assitant Adjunct Professor, QB3

# Outline

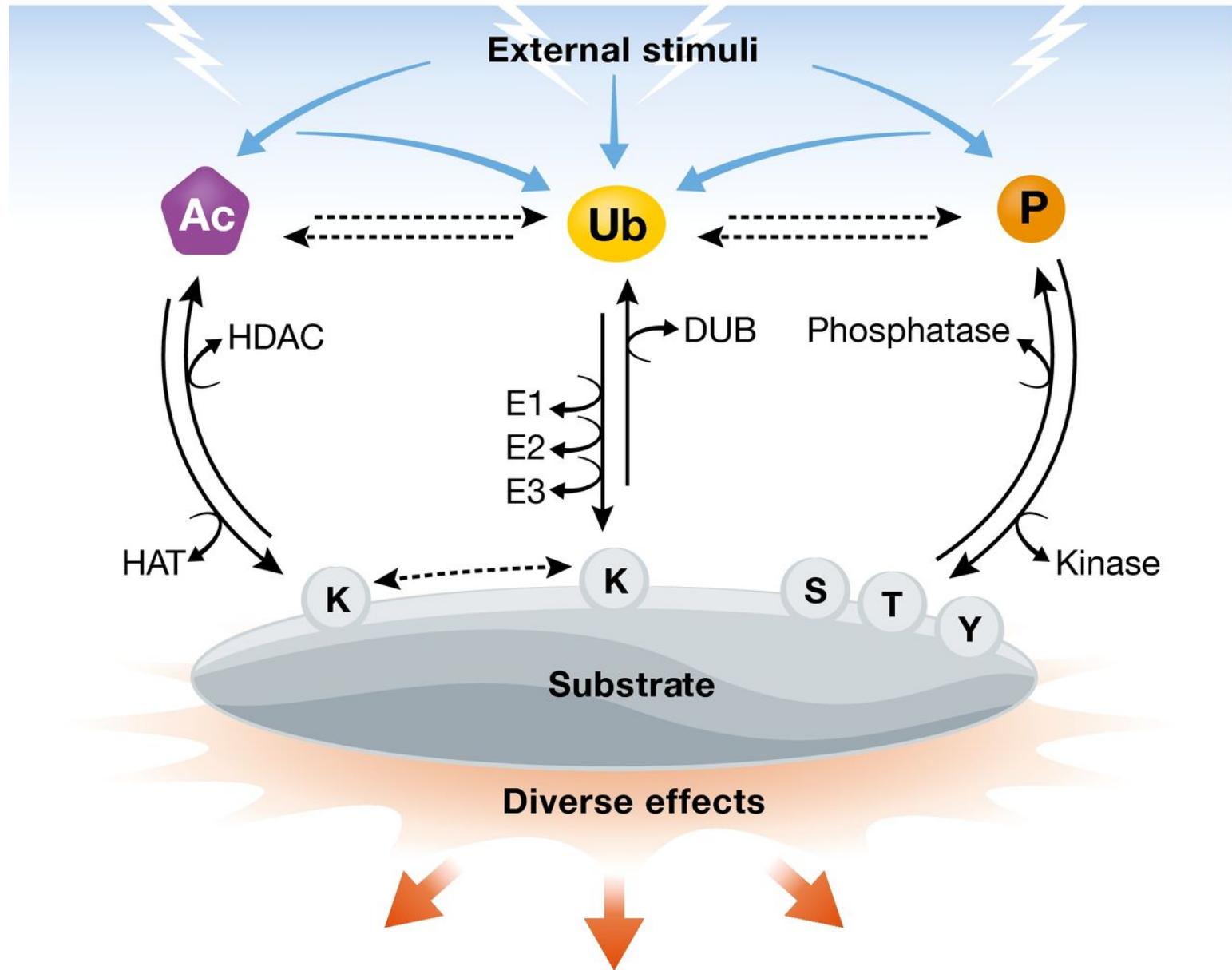
- Rationale for mass spectrometry project
- Approaches to study kinase-substrate interactions
- Introduction to mass spectrometry

# Ubiquitin is a protein post-translational modification with a wide variety of roles

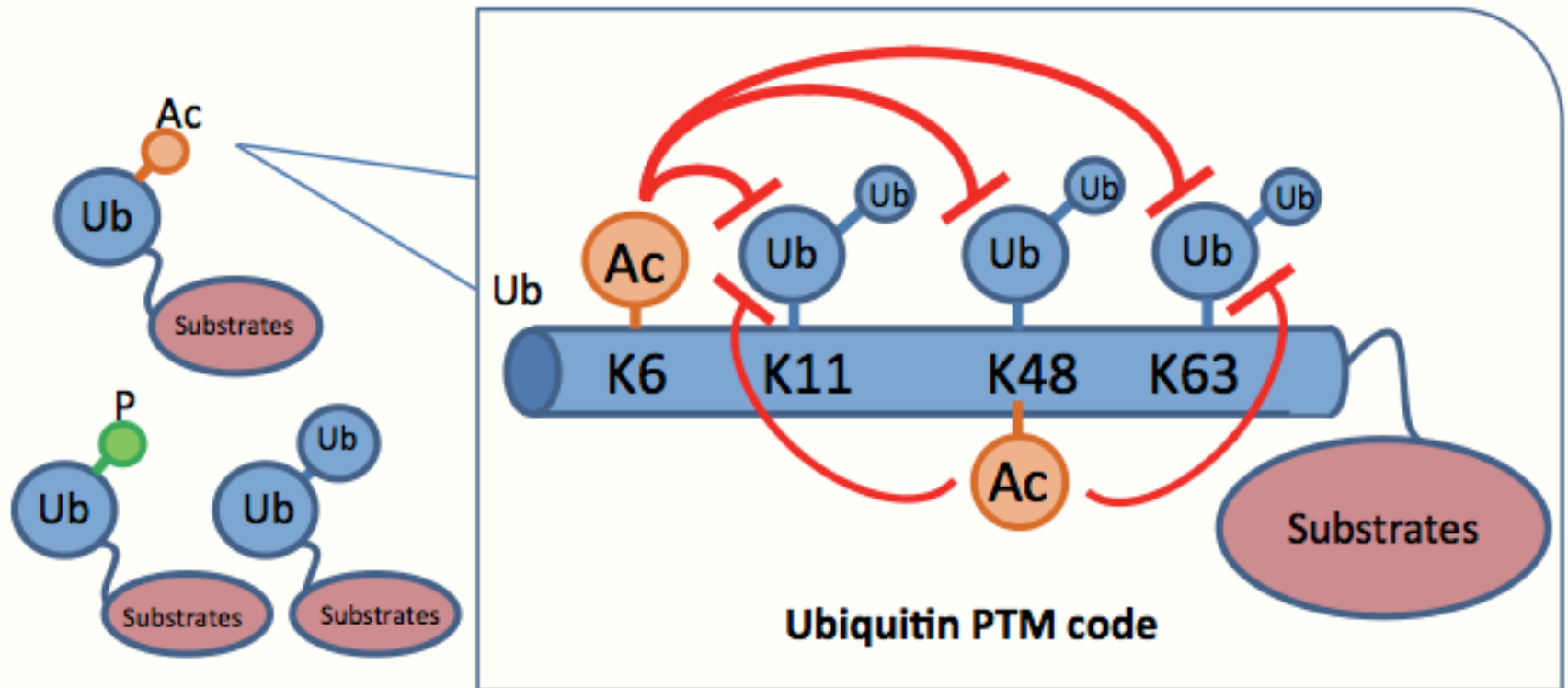




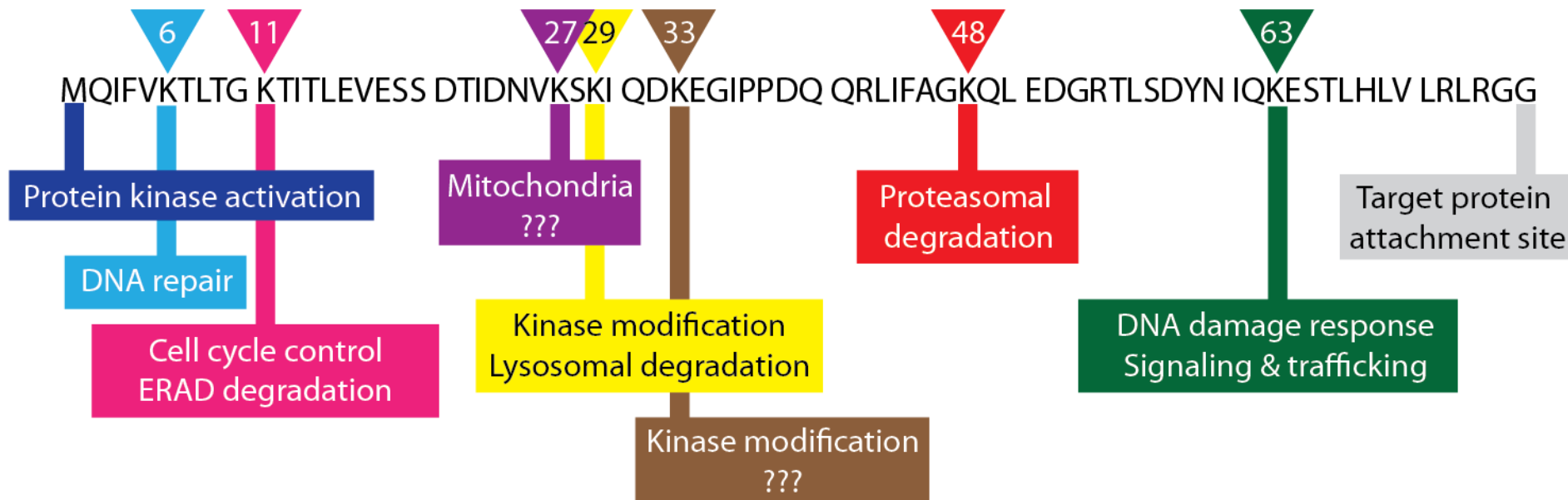
# Ubiquitin itself can be modified by other post-translational modifications



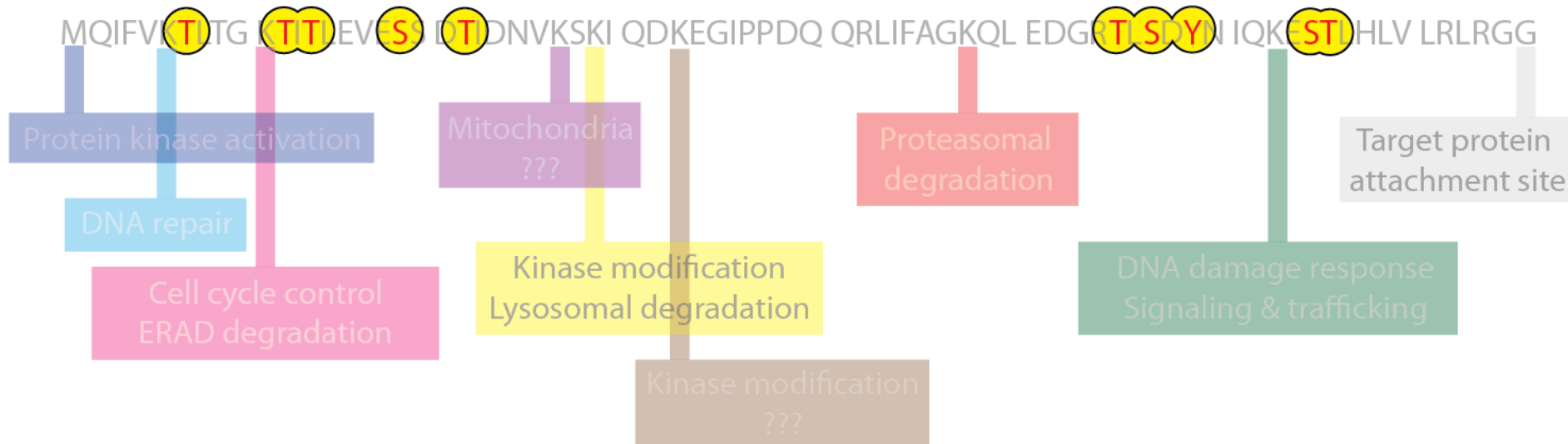
# Lysine acetylation of ubiquitin – blocks ubiquitin chain elongation



# Functions of ubiquitin

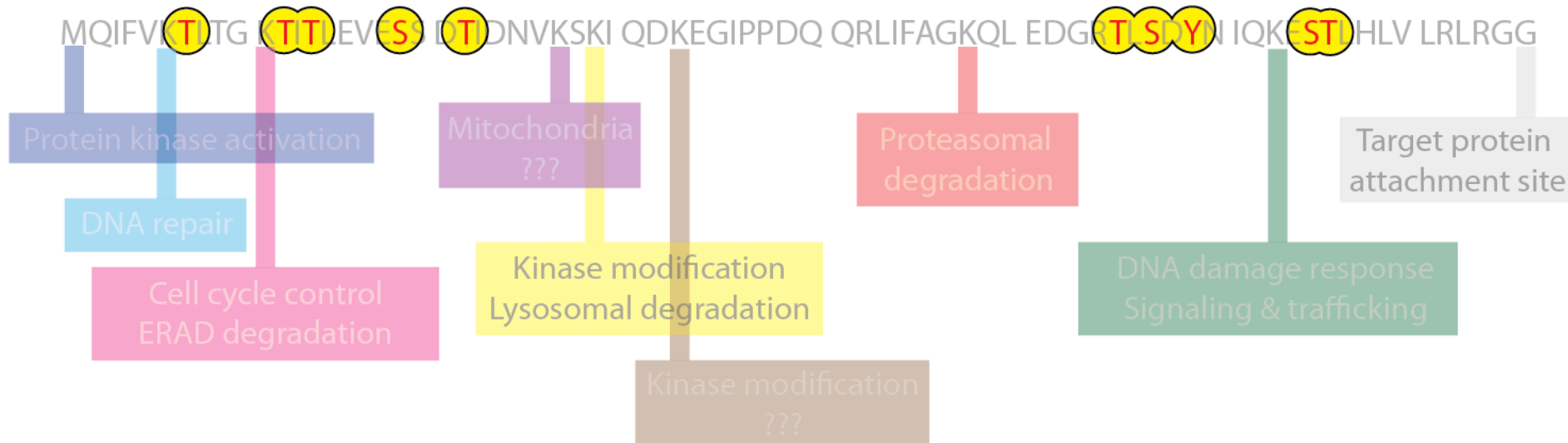


Nearly every S/T/Y on ubiquitin (the most conserved protein) is phosphorylated and conserved from human to yeast



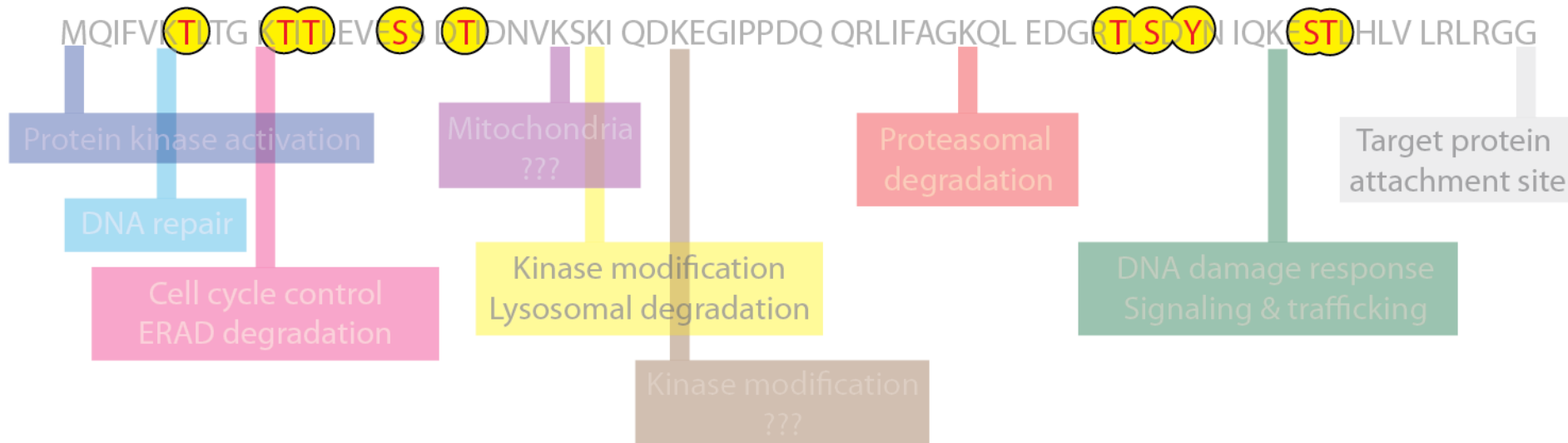
Nearly every S/T/Y on ubiquitin is phosphorylated and conserved from human to yeast

## What kinases phosphorylation ubiquitin?



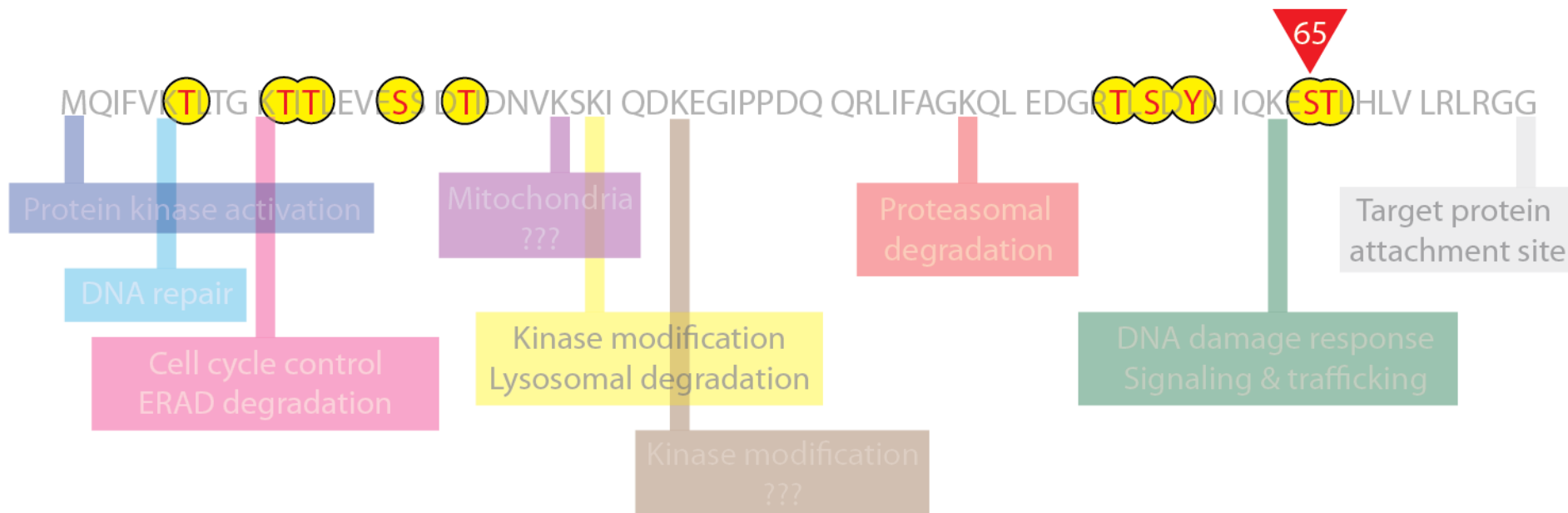
Nearly every S/T/Y on ubiquitin is phosphorylated and conserved from human to yeast

## What kinases phosphorylation ubiquitin?

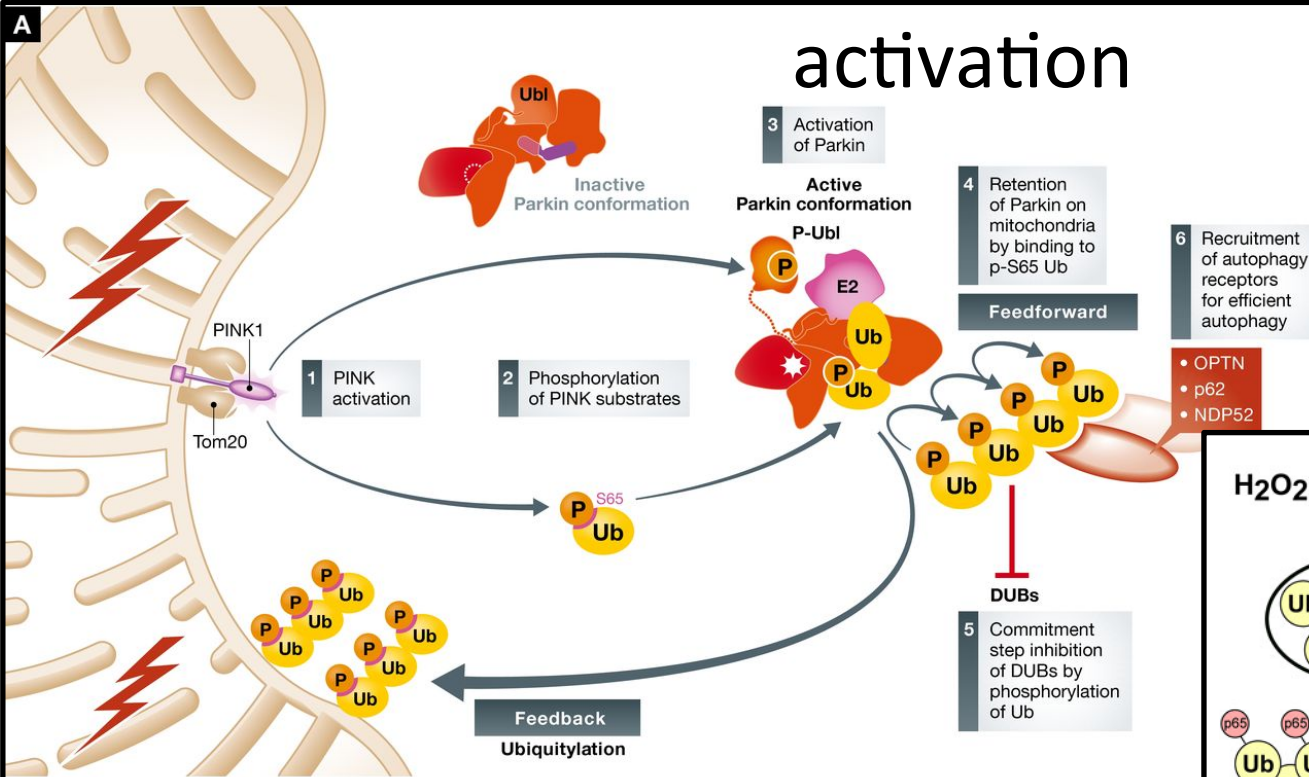


## How does phosphorylation regulate ubiquitin function?

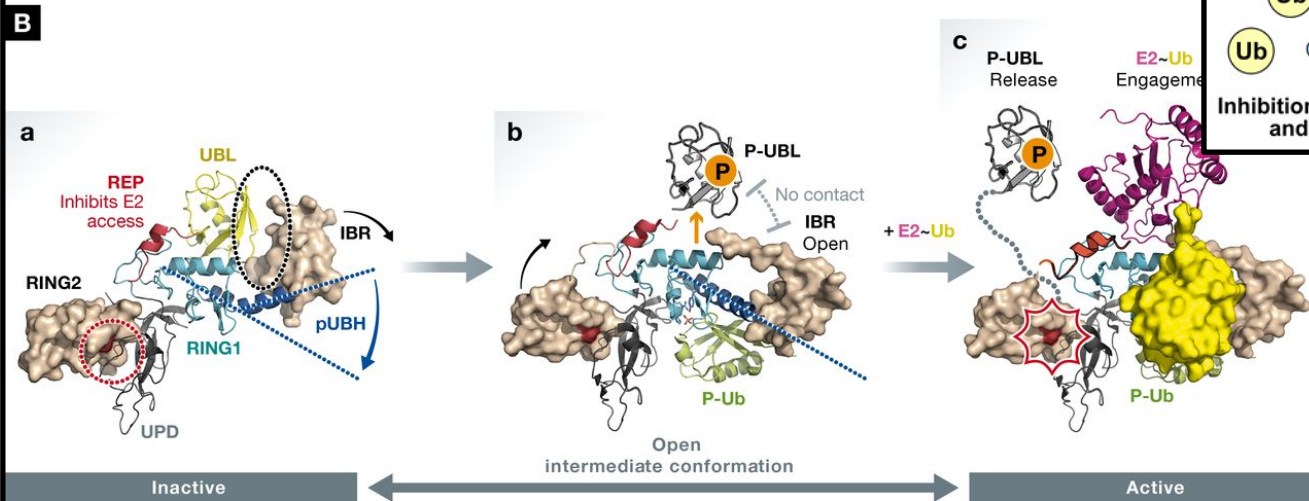
# How does phosphorylation regulate ubiquitin function?



# Human: Pink1 kinase $\rightarrow$ Ub S65p $\rightarrow$ Parkin E3 ligase

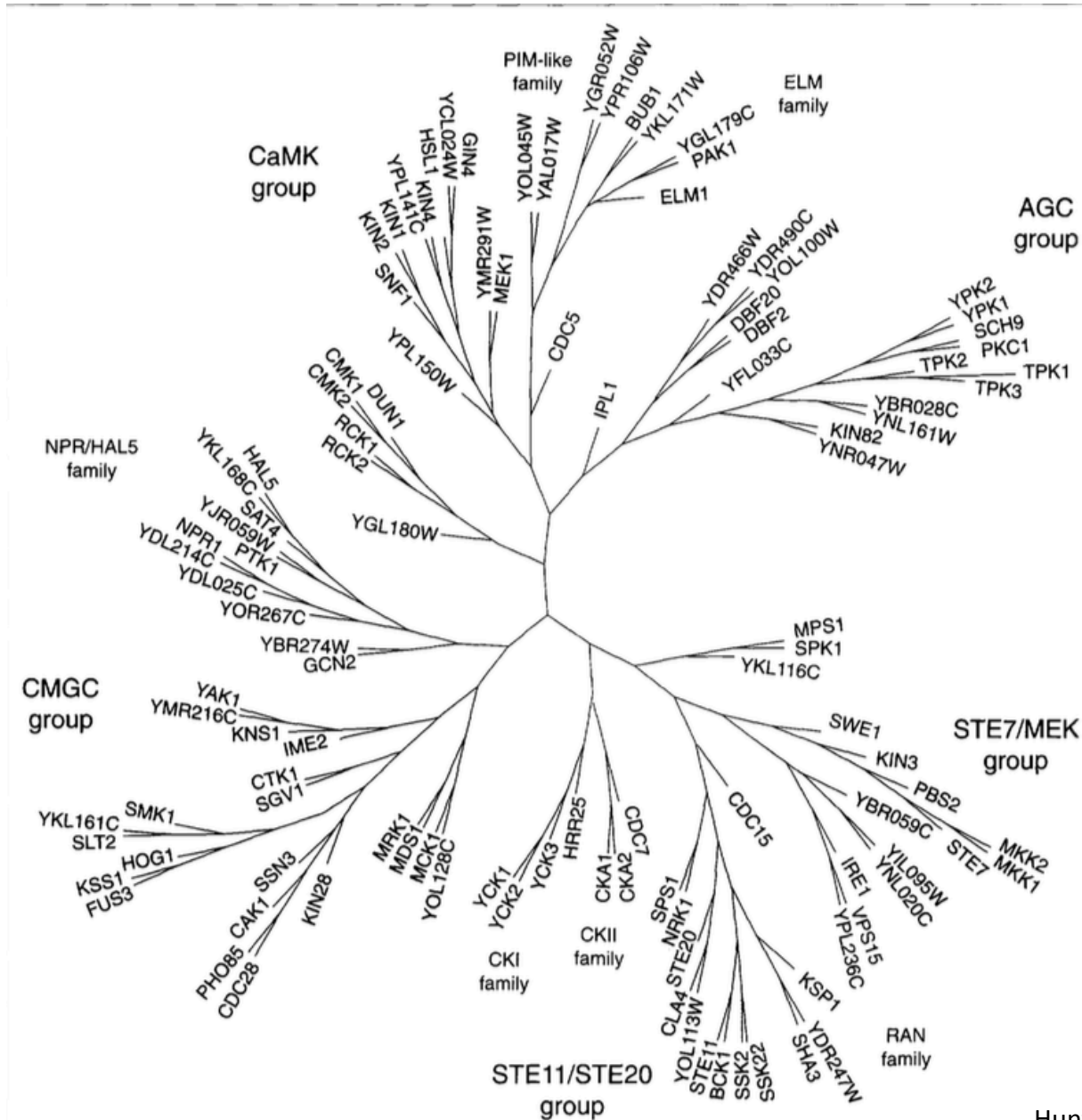


Ub S65p also exists in yeast, despite no Pink1 ortholog. What is the kinase?



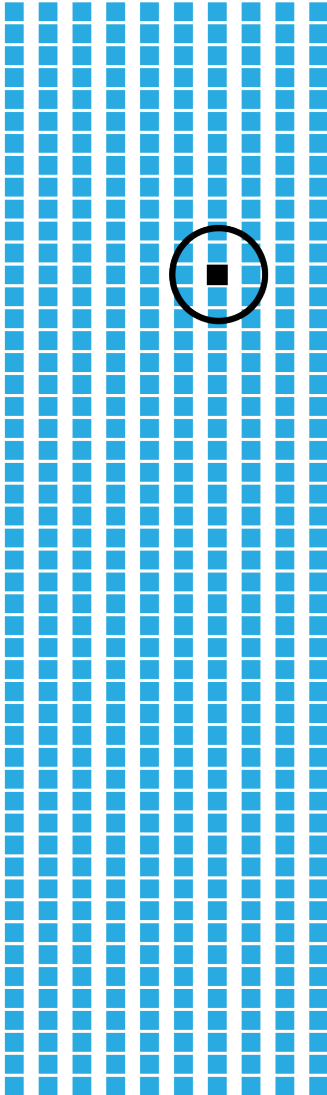


# Approaches to study kinase-substrate interactions

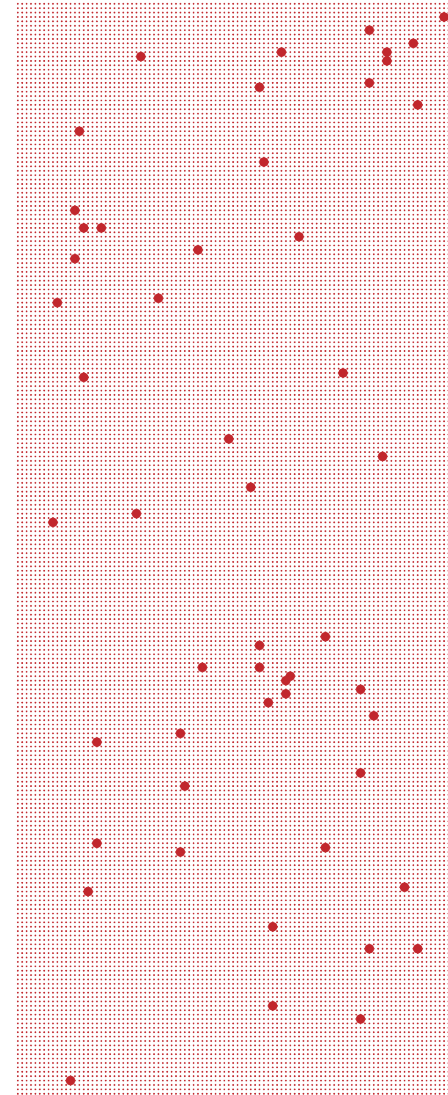


# Connecting enzymes and substrates is challenging

> 500 Protein Kinases



> 25,000 Phosphosites



How can we connect these?



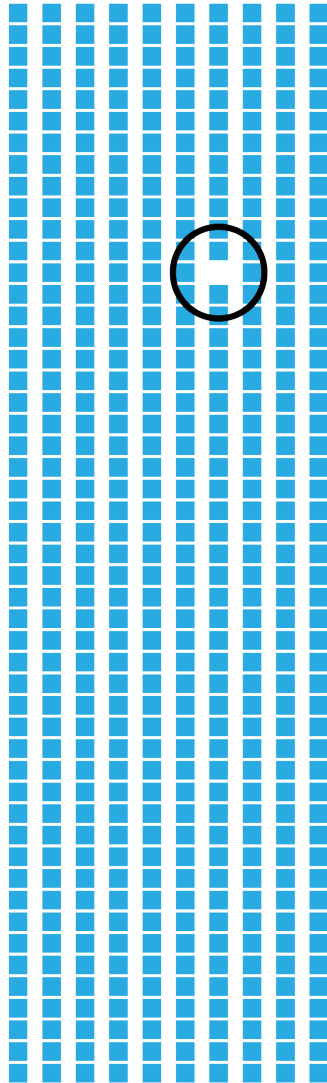
Two lines of evidence are typically required:

1. *in vivo*
  - Overexpression

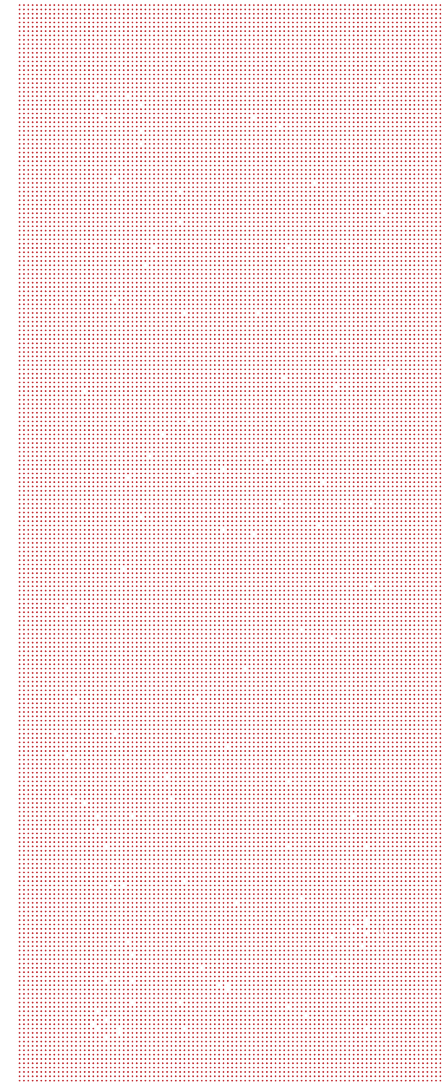


# Connecting enzymes and substrates is challenging

> 500 Protein Kinases



> 25,000 Phosphosites



How can we connect these?



Two lines of evidence are typically required:

1. *in vivo*

- Overexpression
- Knockdown



# Connecting enzymes and substrates is challenging

> 500 Protein Kinases

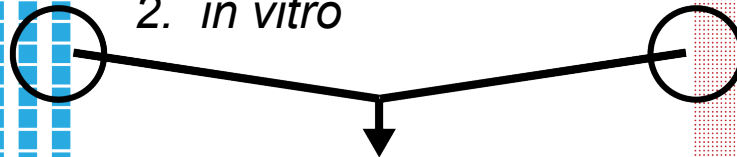
> 25,000 Phosphosites

How can we connect these?

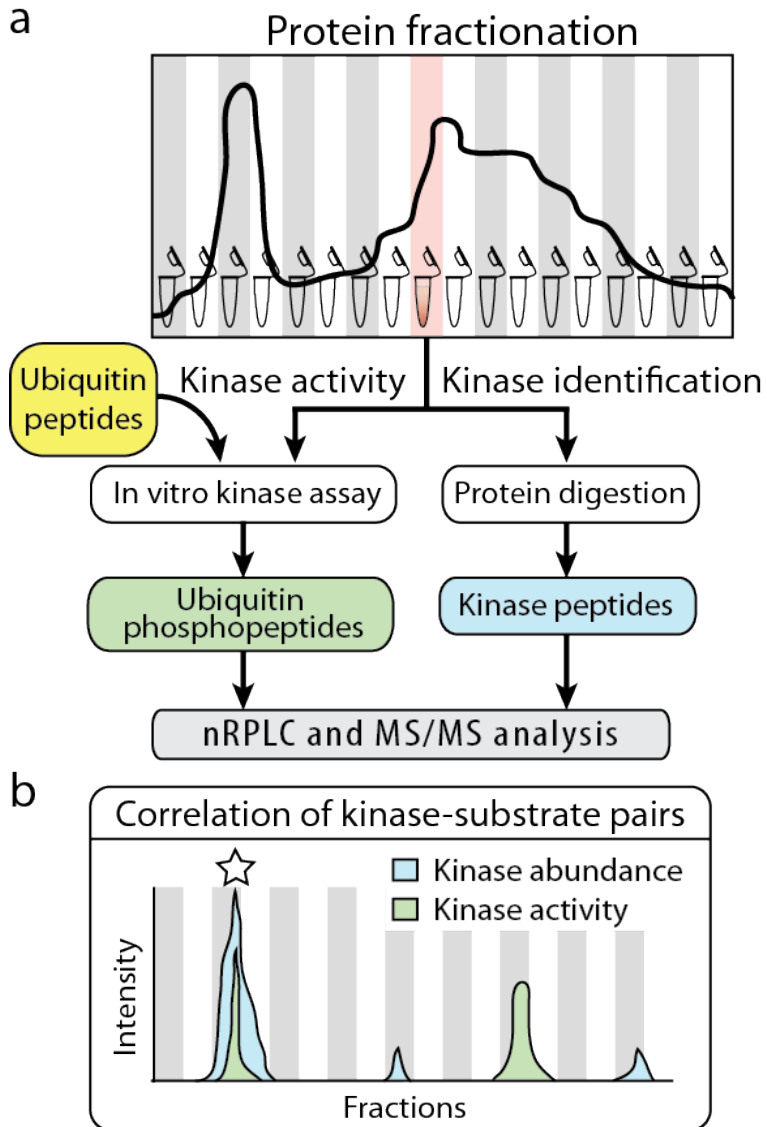


Two lines of evidence  
are typically required:

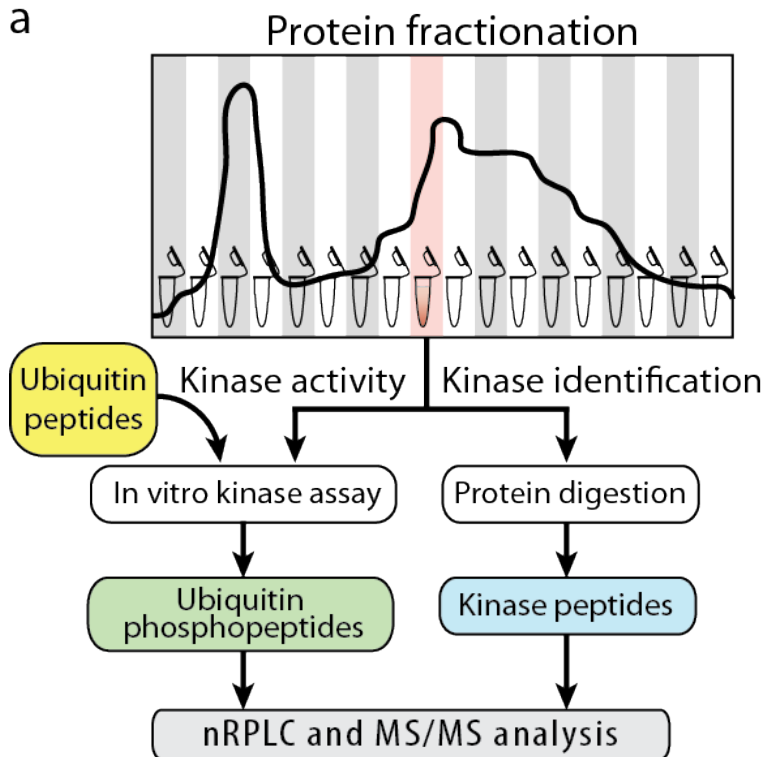
2. *in vitro*



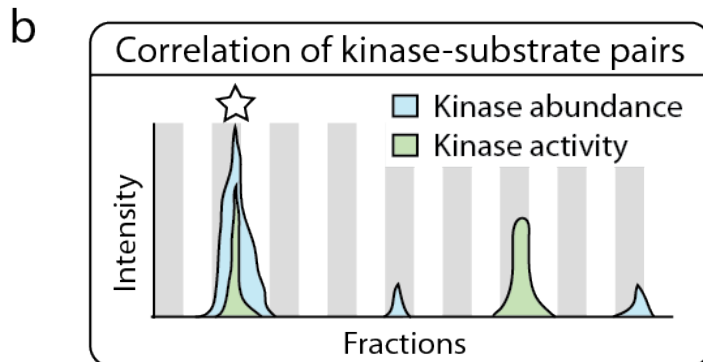
# Kinase activity profiling approach



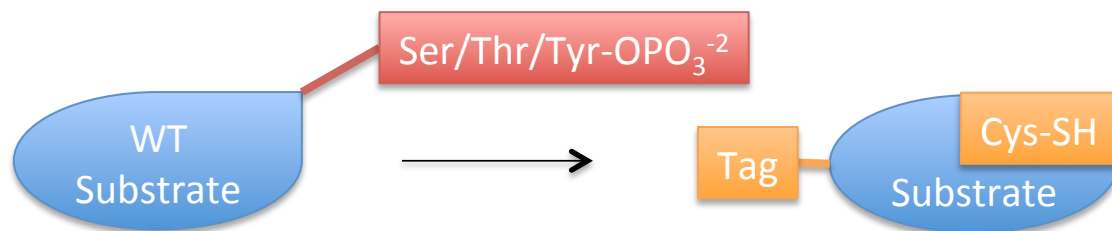
# Kinase activity profiling approach



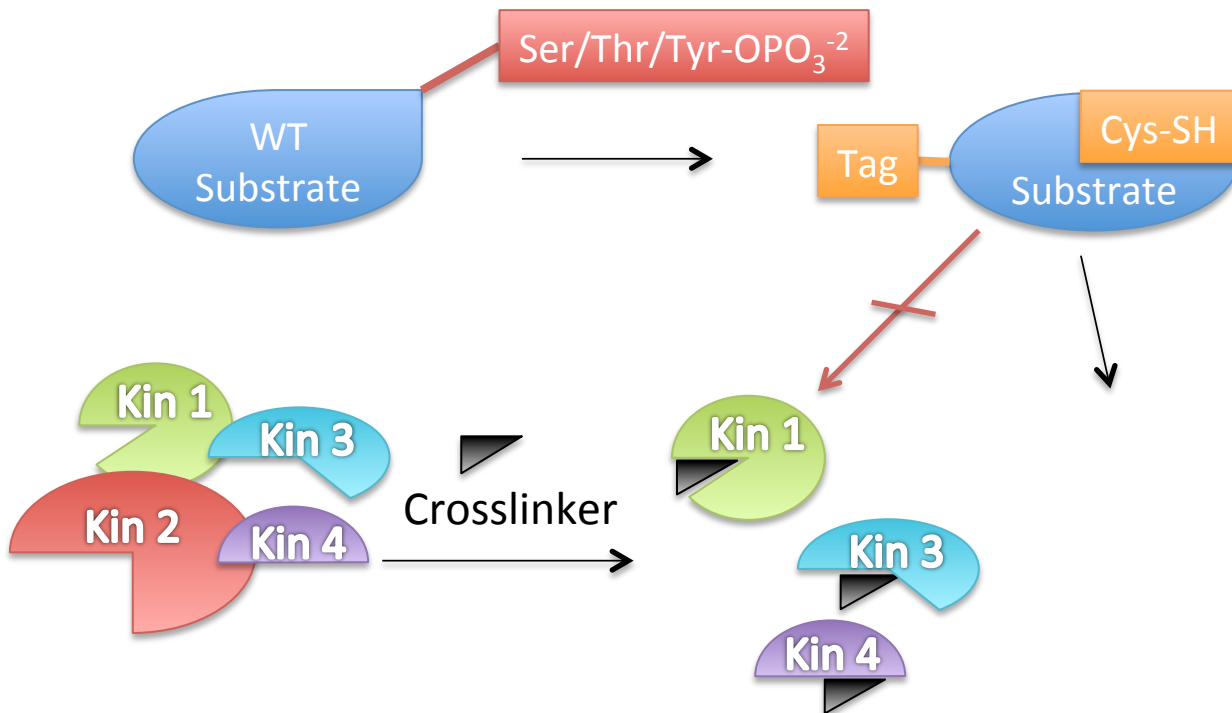
PROS: Un-biased  
CONS: labor intensive, high-false positive



# Chemical Biology approach (Shokat method)

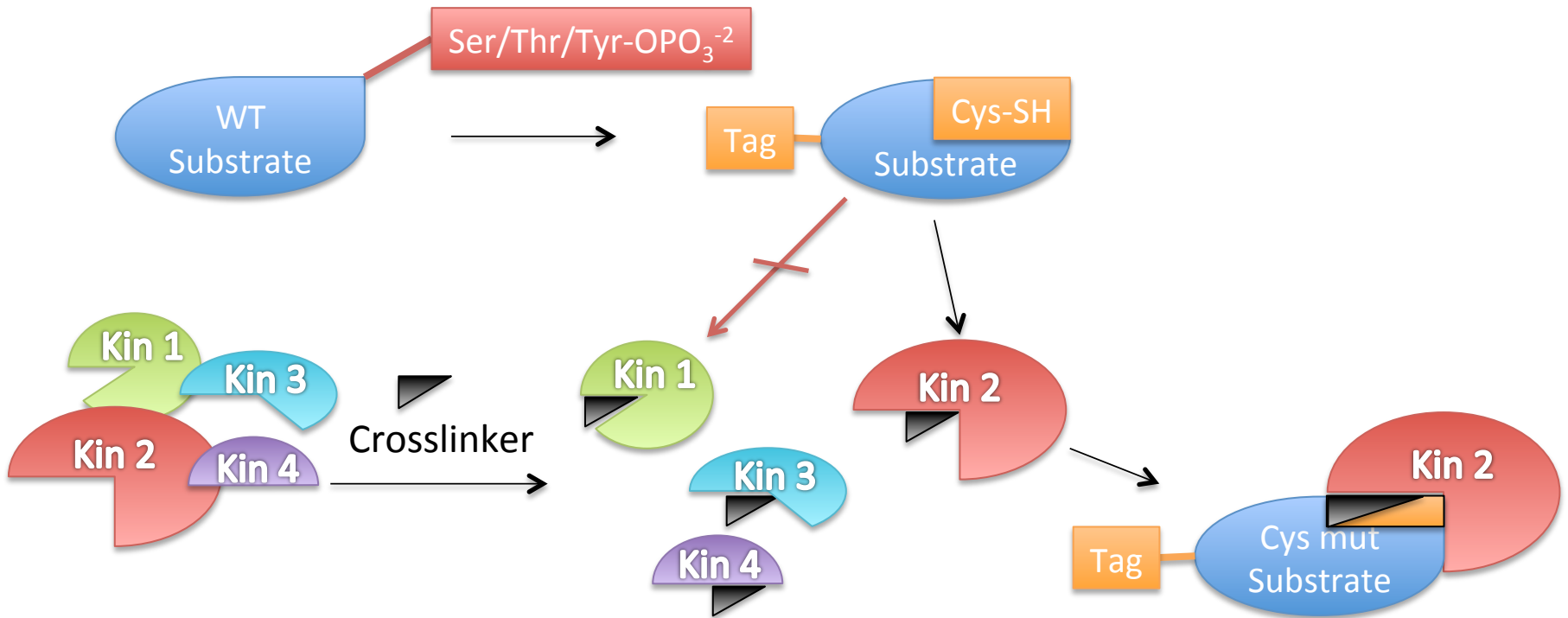


# Chemical Biology approach (Shokat method)

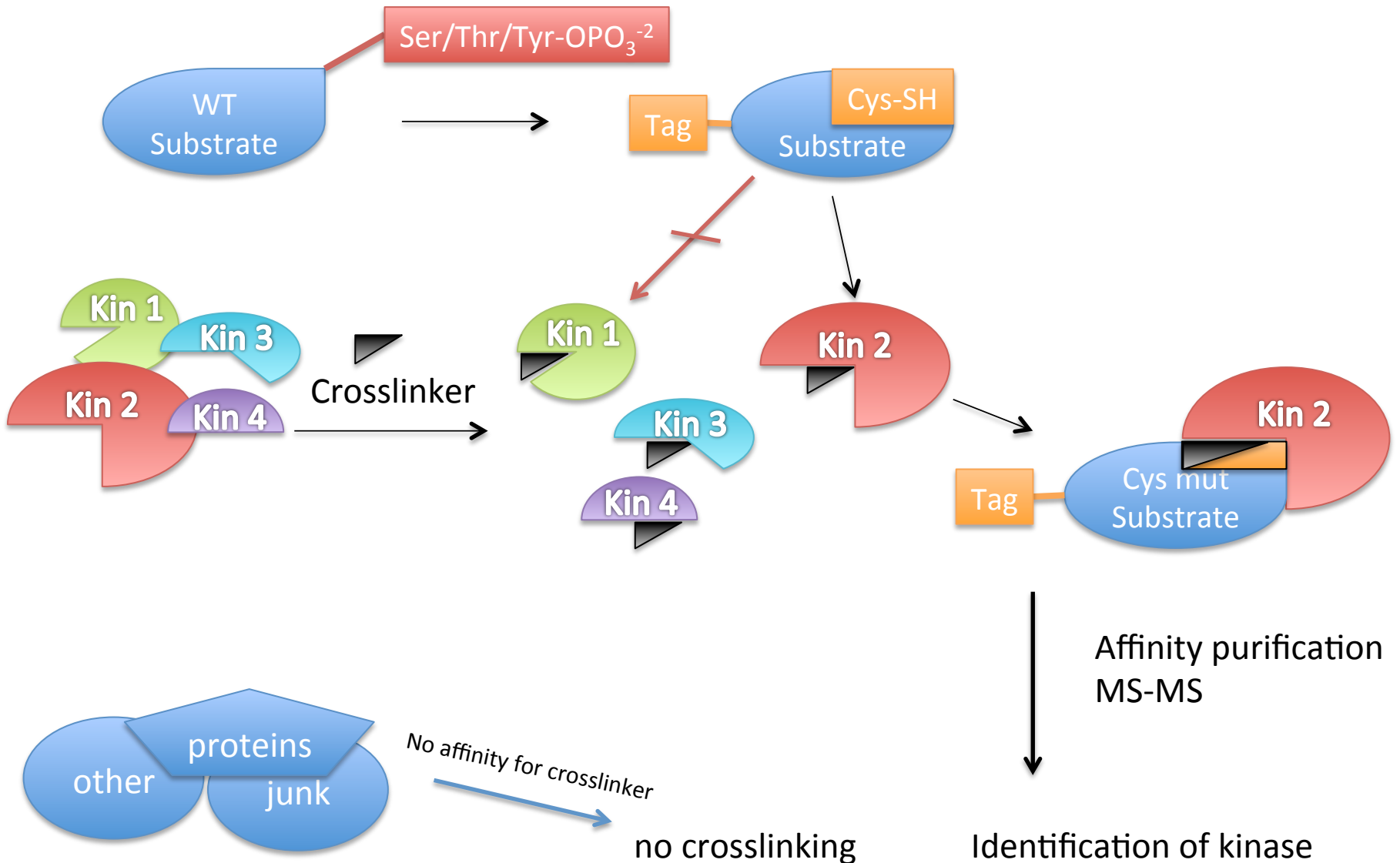




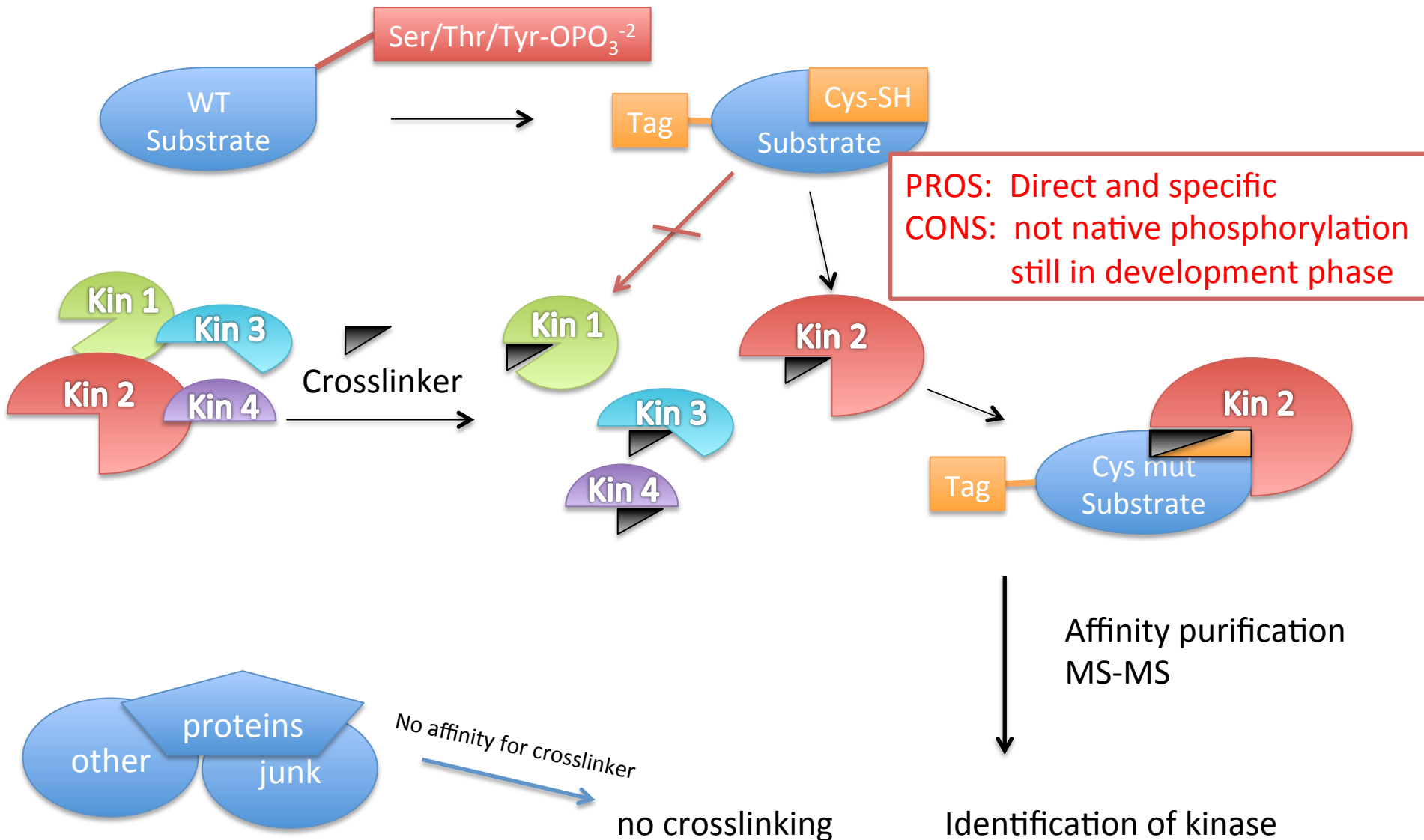
# Chemical Biology approach (Shokat method)



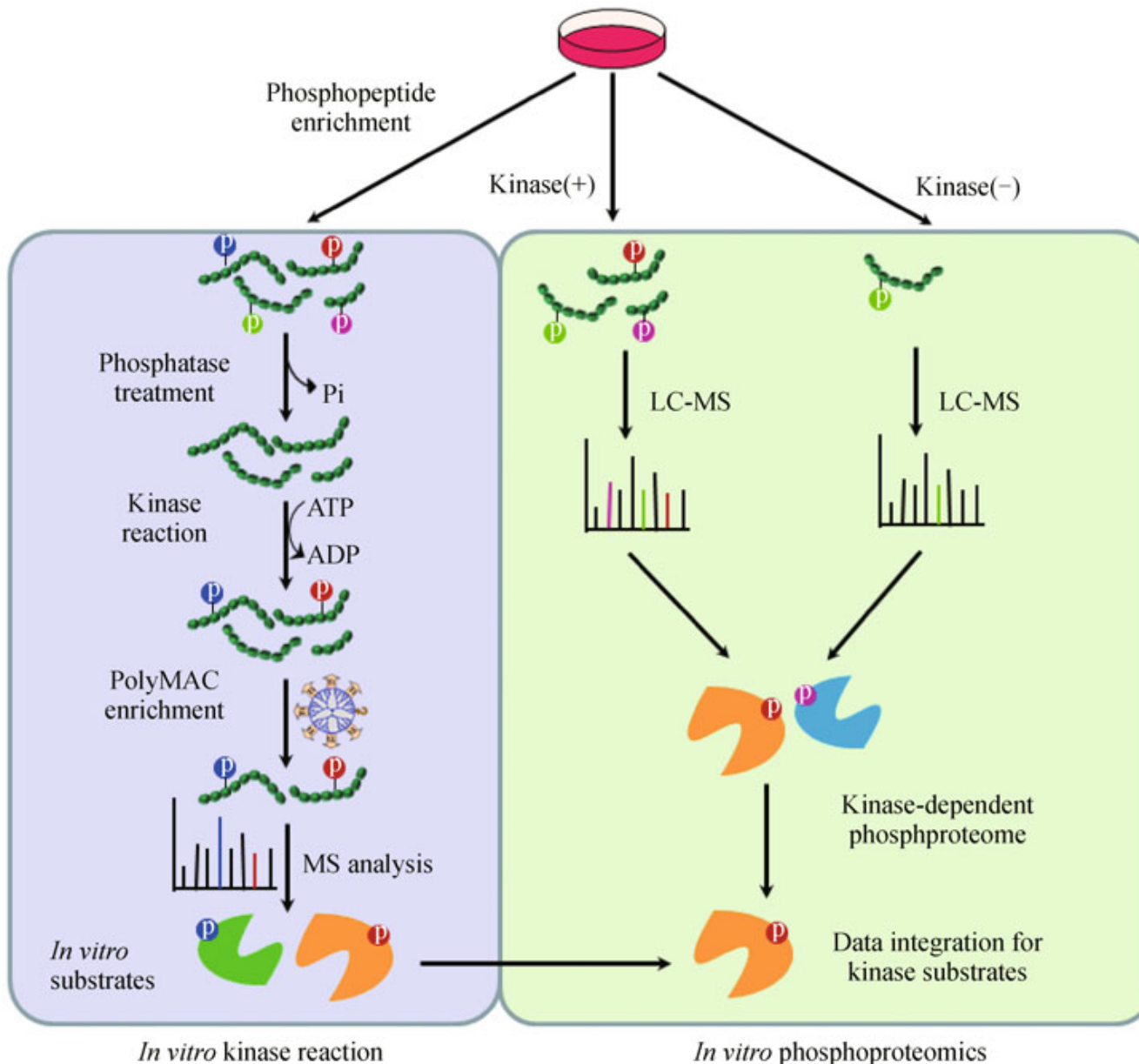
# Chemical Biology approach (Shokat method)



# Chemical Biology approach (Shokat method)

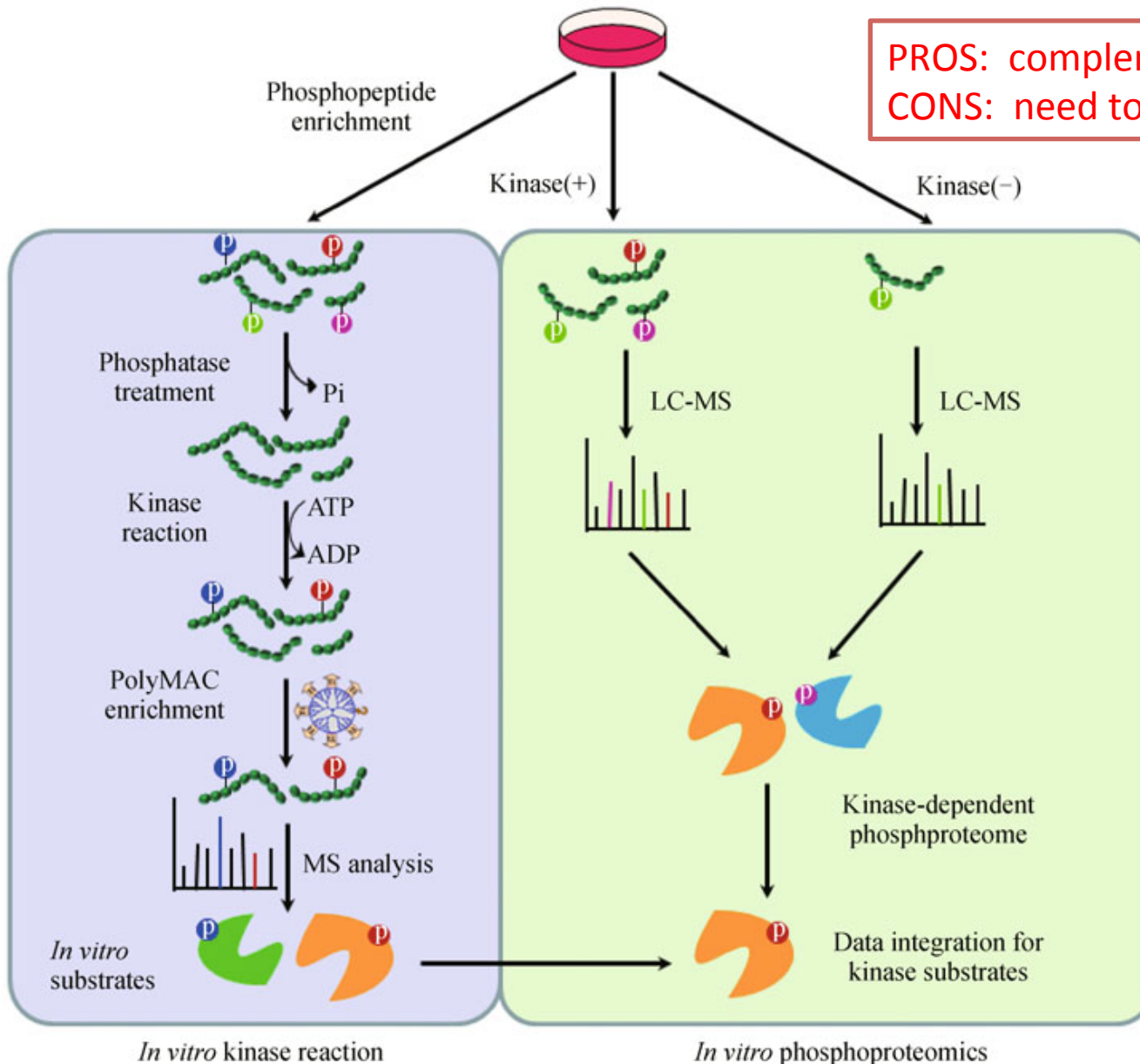


# Kinase directed approaches

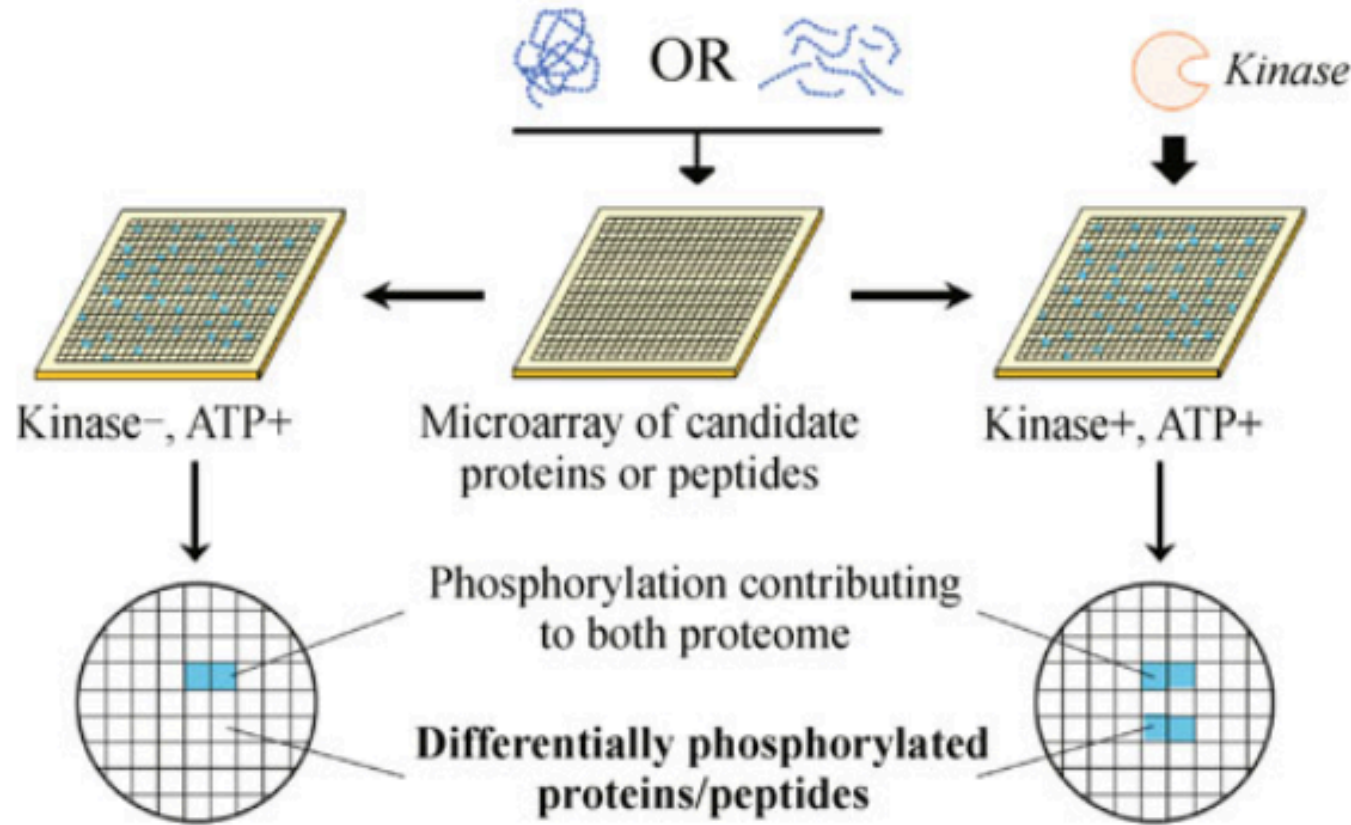


# Kinase directed approaches

PROS: complementary in vivo and in vitro  
CONS: need to know kinase already



# Selection of kinases for this course: protein array approach



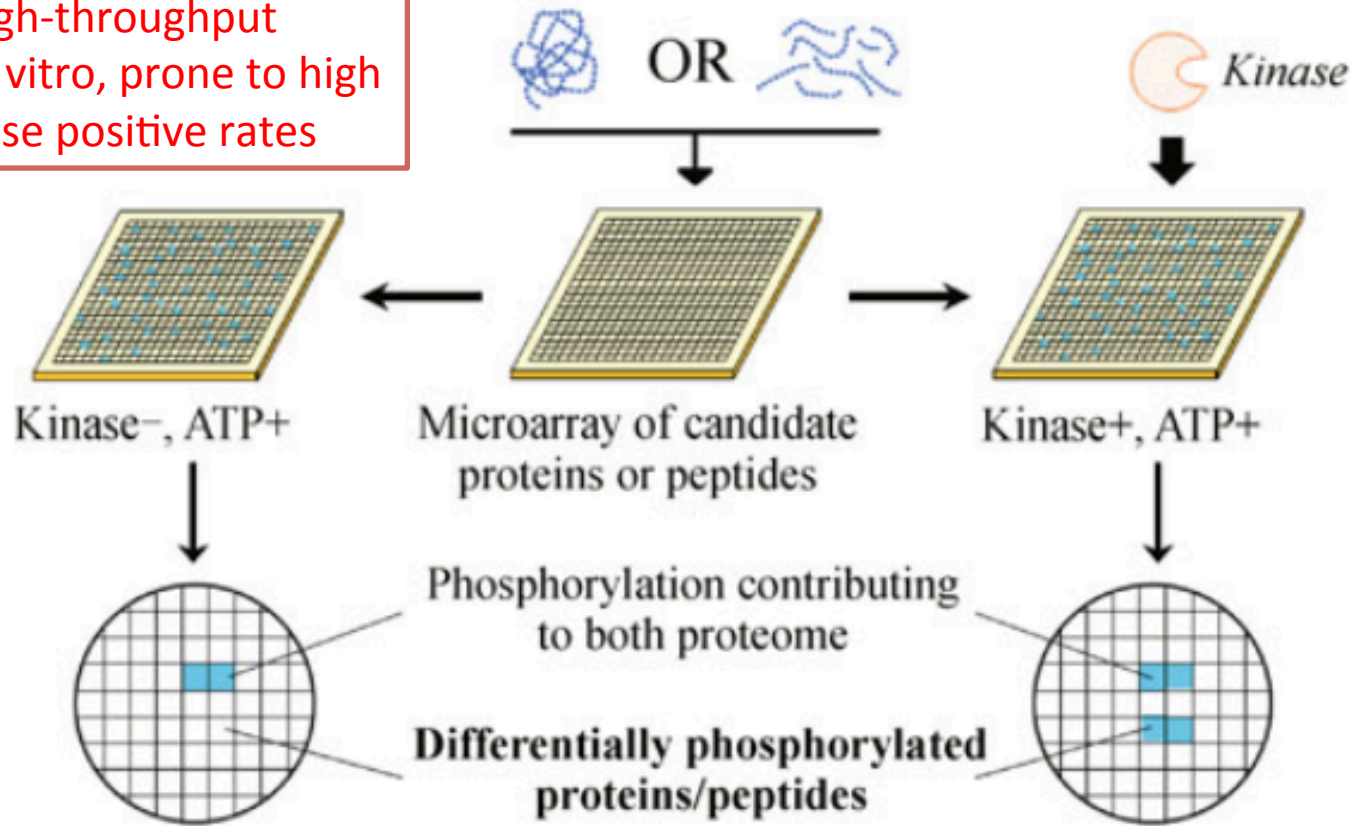
**Figure 2** Kinase assay based on protein array or peptide array. Protein/peptide collections are spotted on the microarray, followed by the incubation with a purified active kinase under the reaction condition. Phosphorylation is detected by various methods.





# Selection of kinases for this course: protein array approach

PROS: High-throughput  
CONS: In vitro, prone to high false positive rates



**Figure 2** Kinase assay based on protein array or peptide array. Protein/peptide collections are spotted on the microarray, followed by the incubation with a purified active kinase under the reaction condition. Phosphorylation is detected by various methods.



**In short, well-established methods to map kinase-substrate relationship require one of the following:**

**(1) Prior knowledge of kinase  
→ substrate hunting**

**(2) A labor intensive brute force approach  
→ kinase hunting**

**(3) Serendipitous luck**

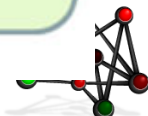
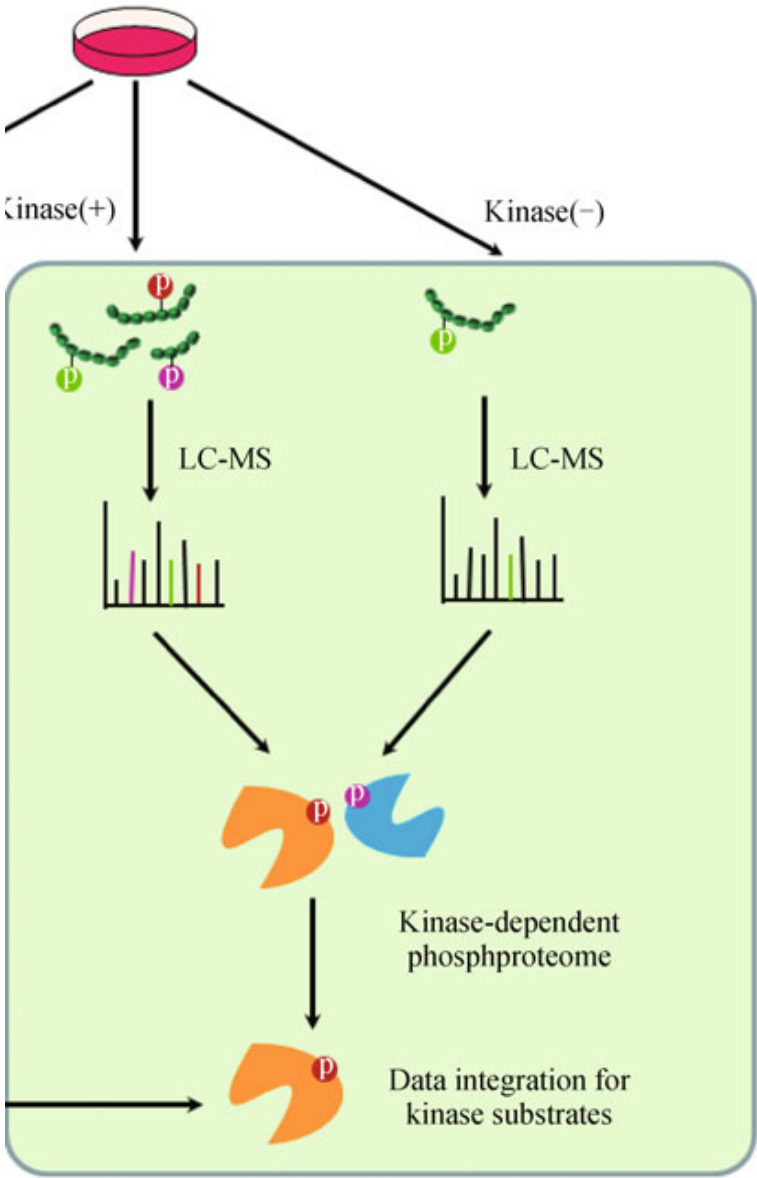
**\*\*\*note other approaches do exist: phage display, yeast 2-hybrid, genetic interaction. But they all suffer from one of the primary CONS listed for methods described here.**



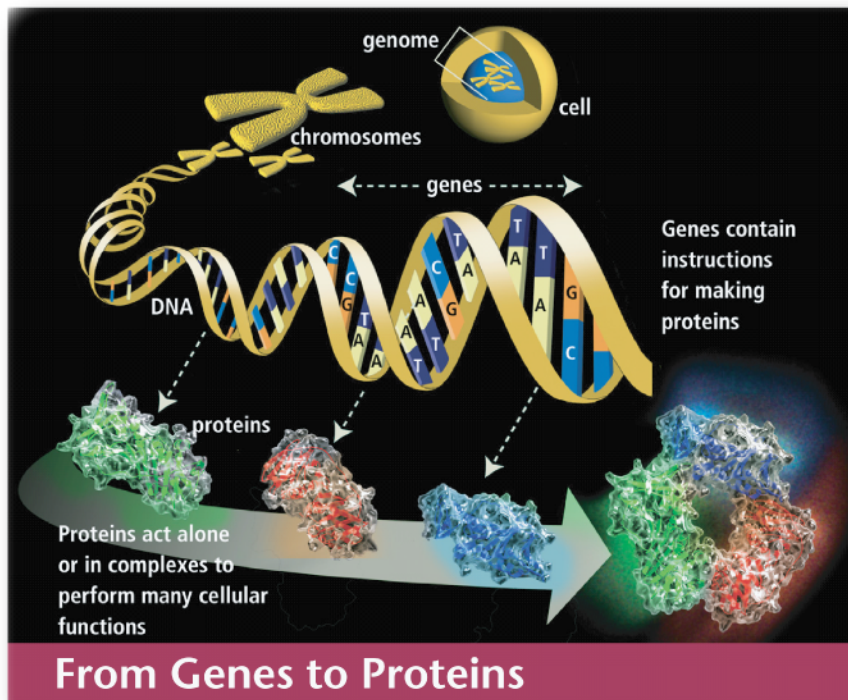


# Selection of kinases for this course: protein array results directing a kinase-directed approach

Paper	Human Kinase	Yeast kinase?
Human kinase array	NEK2	KIN3
Human kinase array and YEAST kinase array	WEE1	SWE1
Yeast kinase array		CMK1
Yeast kinase array		TPK1
Yeast kinase array		ALK1
Yeast kinase array (Youle autophagy paper connected Pink1, phosphoubiquitin, and autophagy signaling)		ATG1



# From genes to proteins



[www.doegenomes.org](http://www.doegenomes.org)

- The human genome codes ~ 25,000 proteins after modification > **500,000**
  - the human body consists of  $10^{14}$  cells
  - each cell makes ~ 15,000 different proteins

- what are they?
- where are they?
- how many copies are present?
- what is their function?
- when are they made?
- what proteins do they interact with?
- how are they modified?

# Mass spectrometry based proteomics

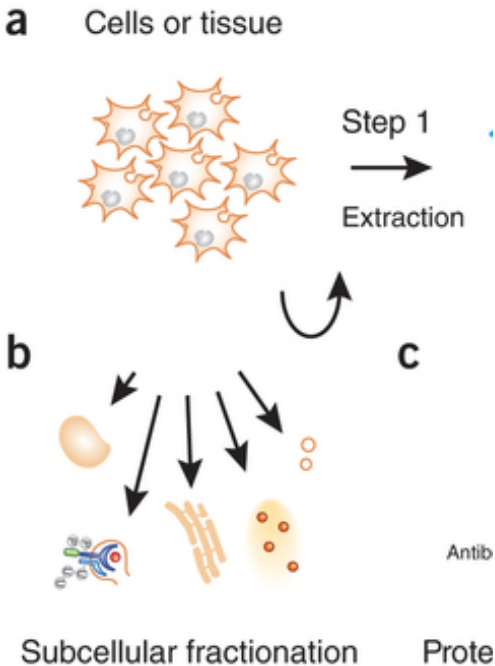


Velos Orbitrap



Orbitrap Fusion

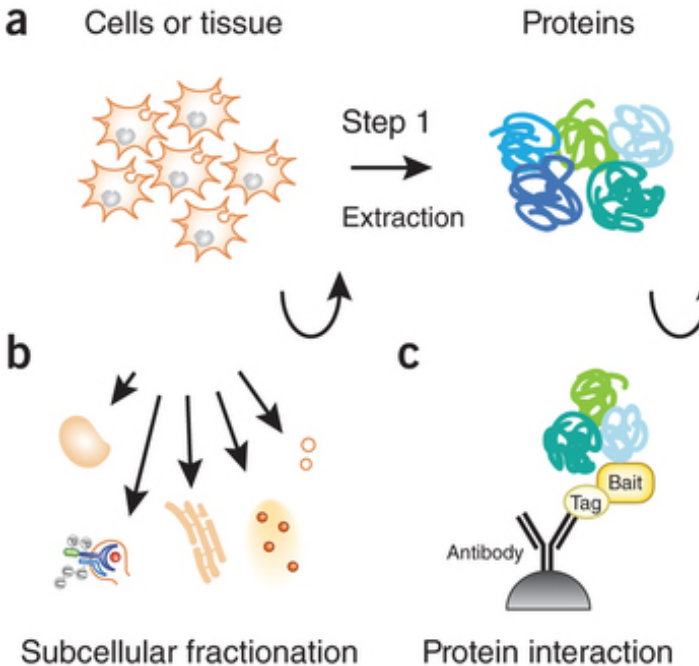
# Proteomics Workflow



## Considerations:

- Qualitative: what proteins are there?
- Quantitative: What differences in proteins or PTMs between conditions?
  - Different cell types
  - Kinase KO
  - Chemical perturbation
  - Etc.

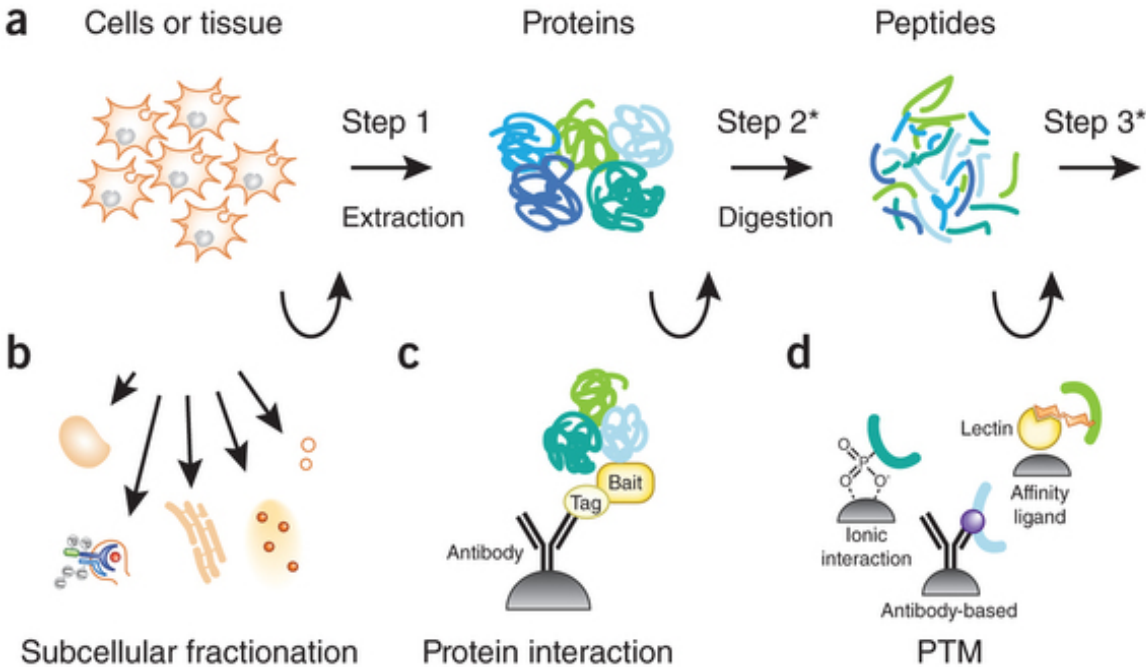
# Proteomics Workflow



## Considerations:

- Native or denaturing?
- Protein purification
- PTM stability

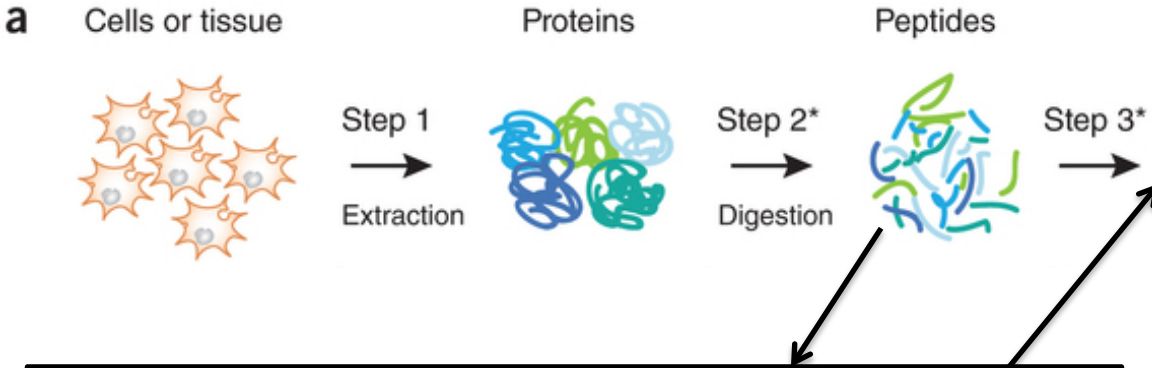
# Proteomics Workflow



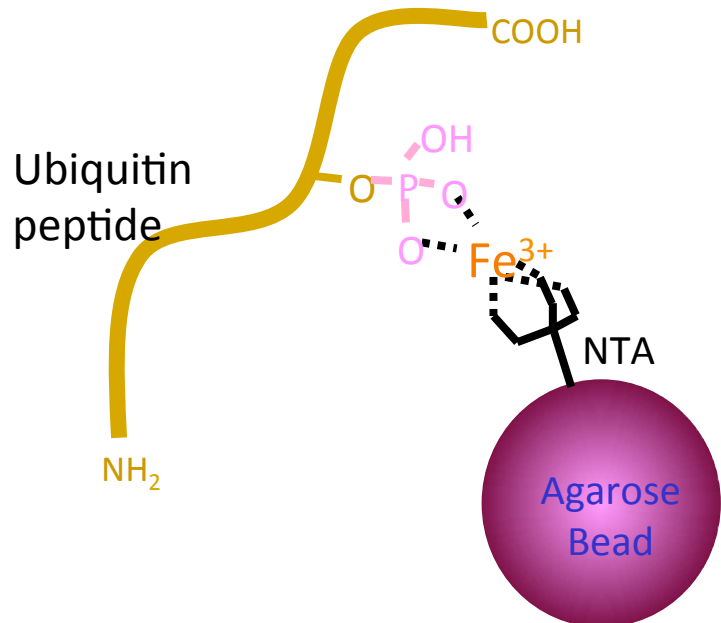
Considerations:

- PTM purification
- Fractionation to reduce complexity

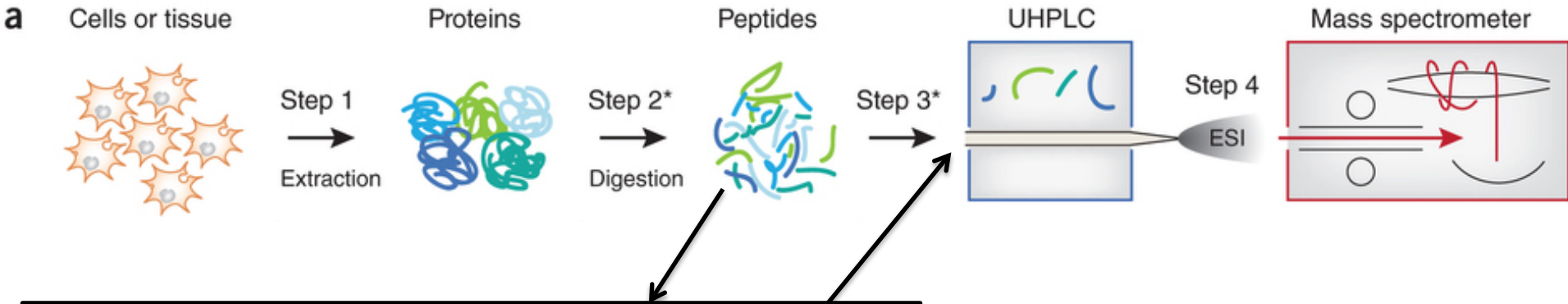
# Proteomics Workflow



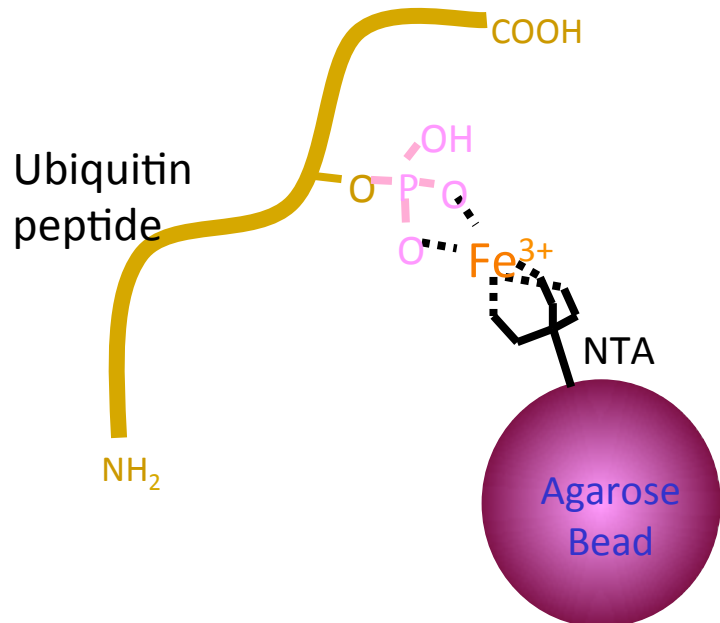
Phosphopeptide enrichment  
Immobilized Metal Affinity Chromatography (IMAC)



# Proteomics Workflow

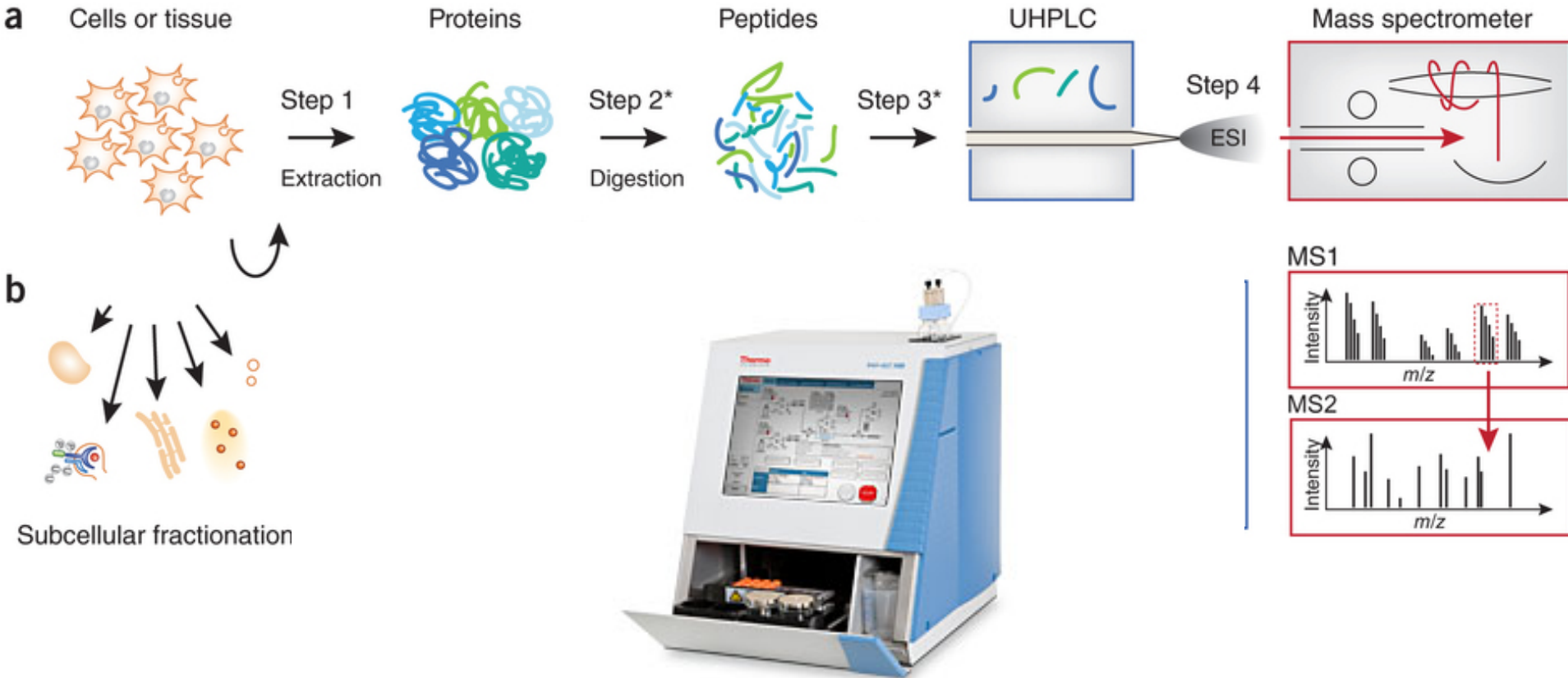


Phosphopeptide enrichment  
Immobilized Metal Affinity Chromatography (IMAC)





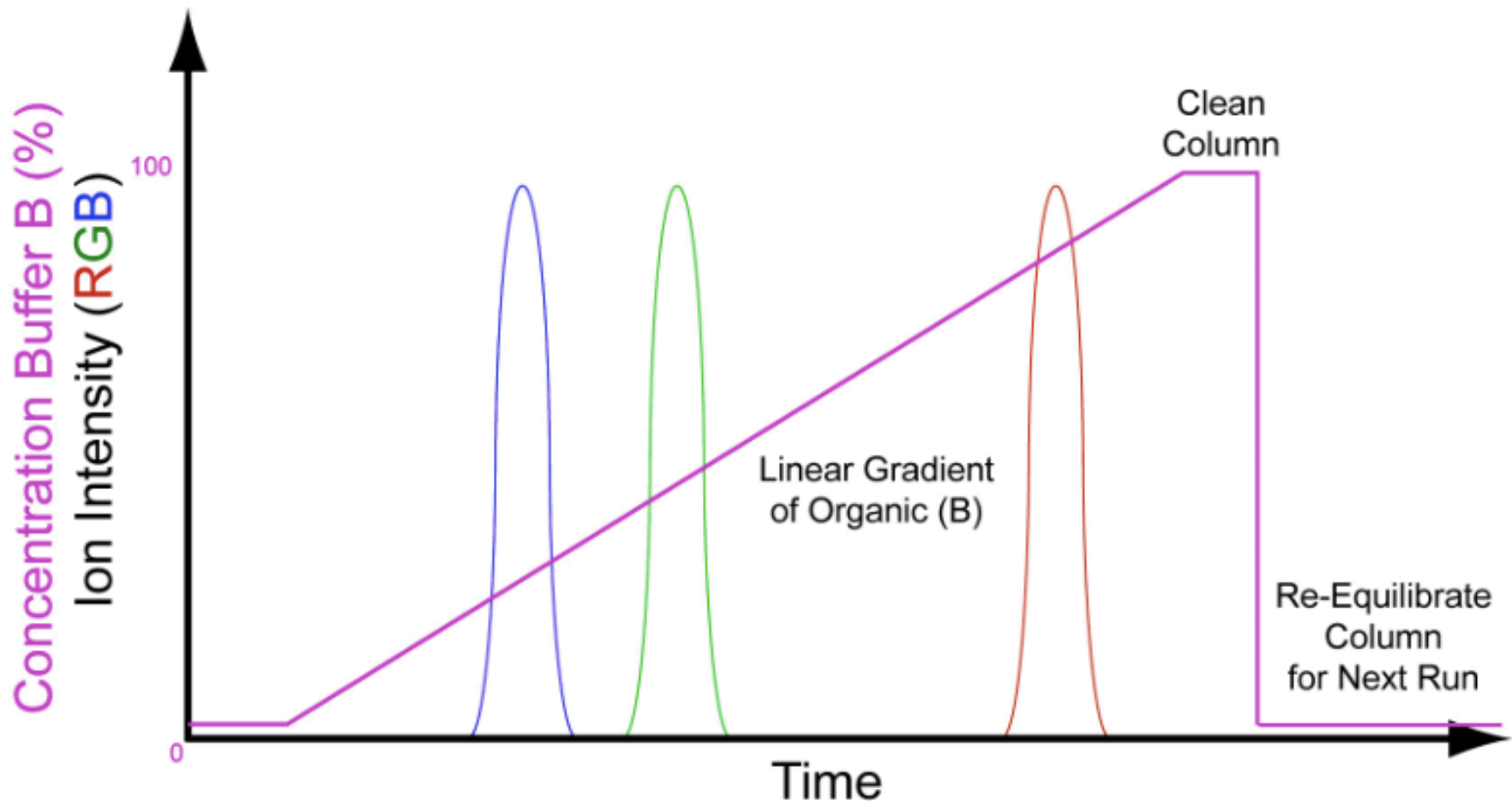
# Proteomics Workflow



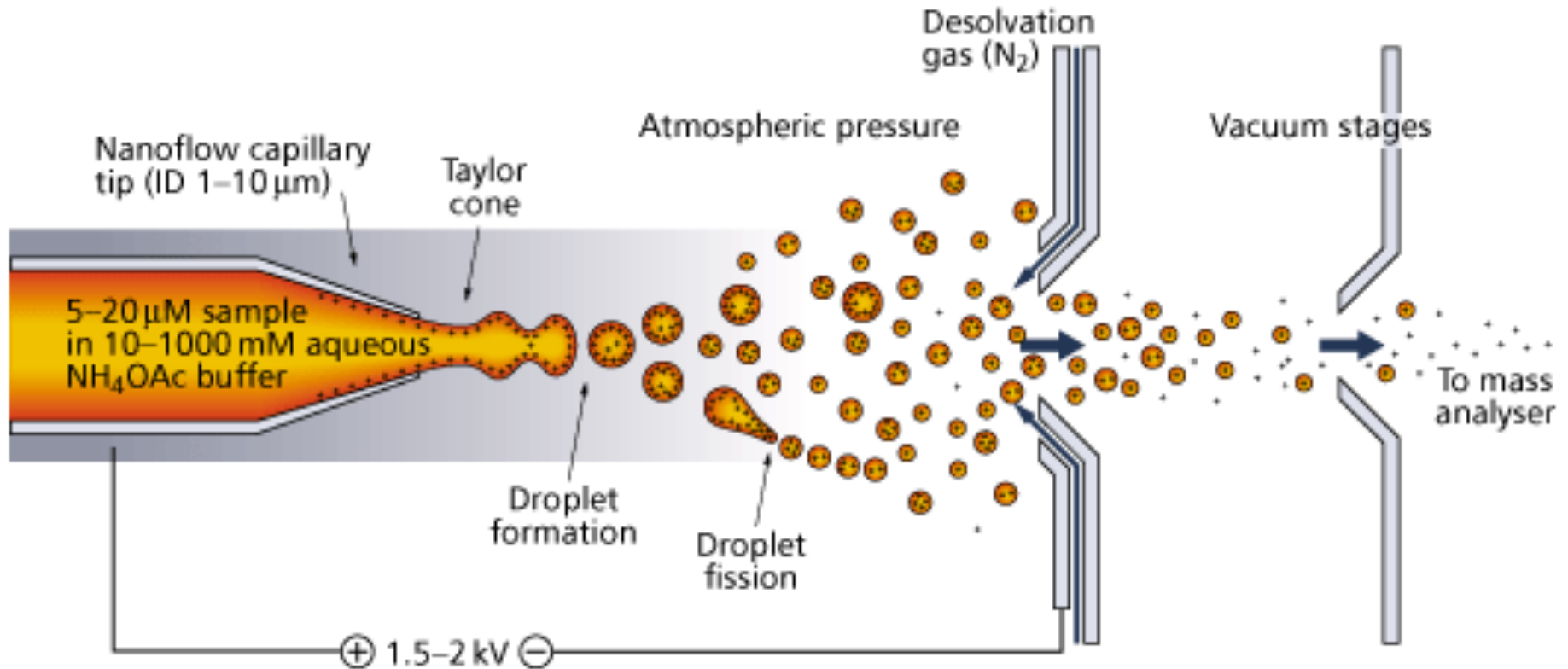
# Reversed-Phase HPLC

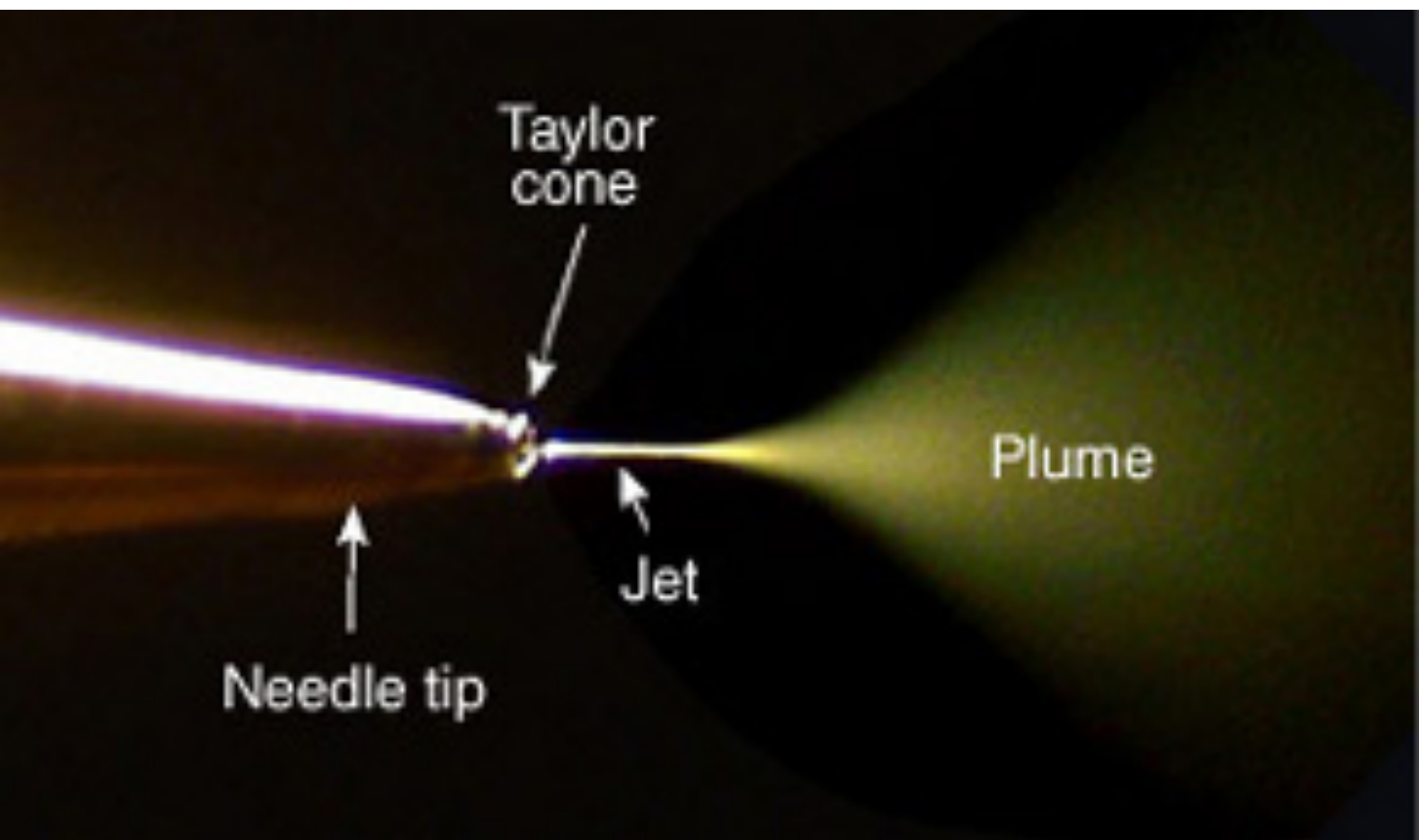
Load peptides: 100% water, 0.1% formic acid

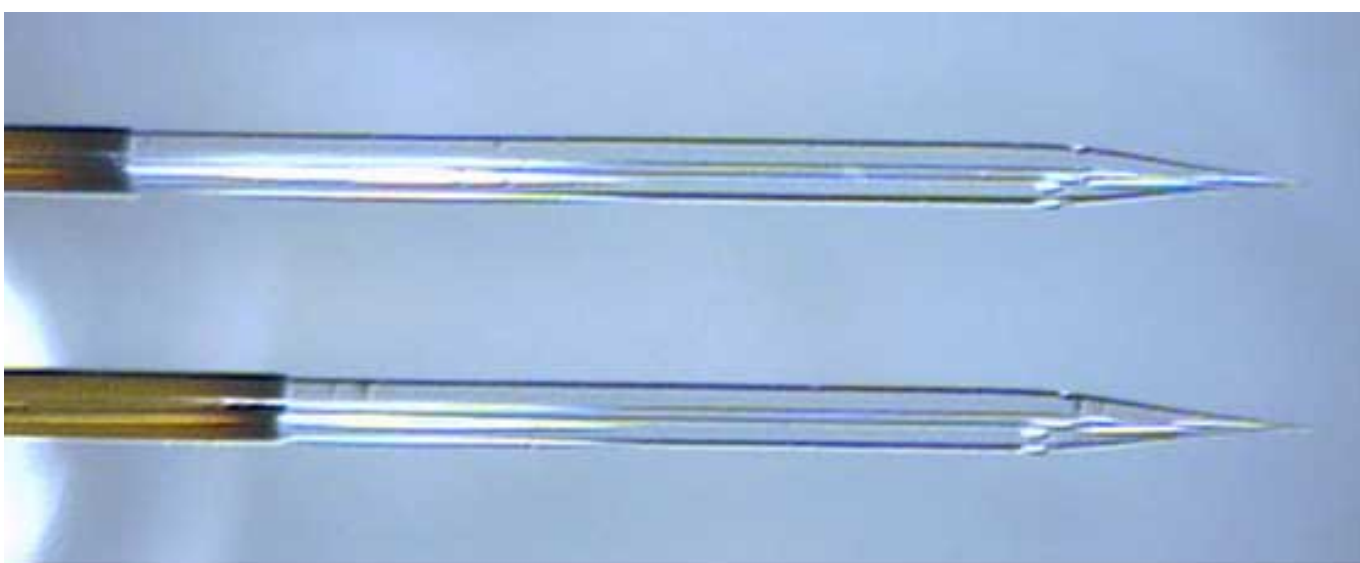
Elute with increasing gradient of acetonitrile, 0.1% formic acid



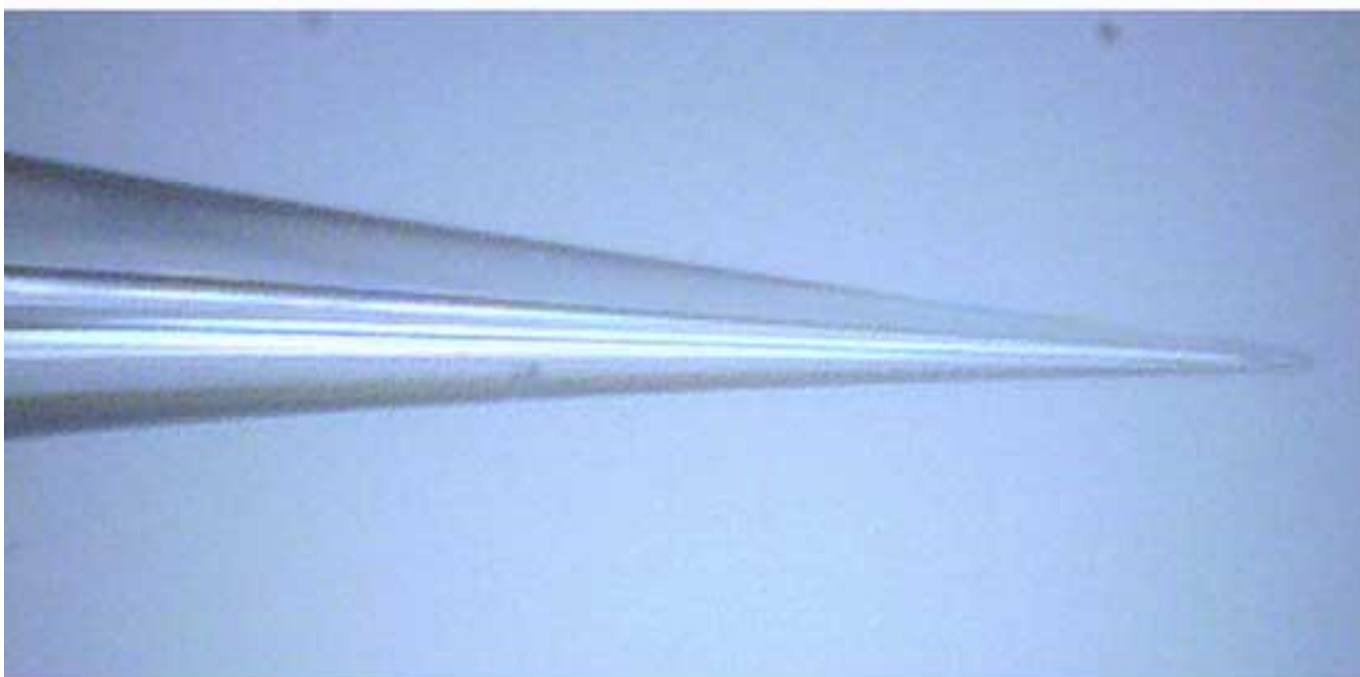
# Getting your sample into mass spec – electrospray ionization





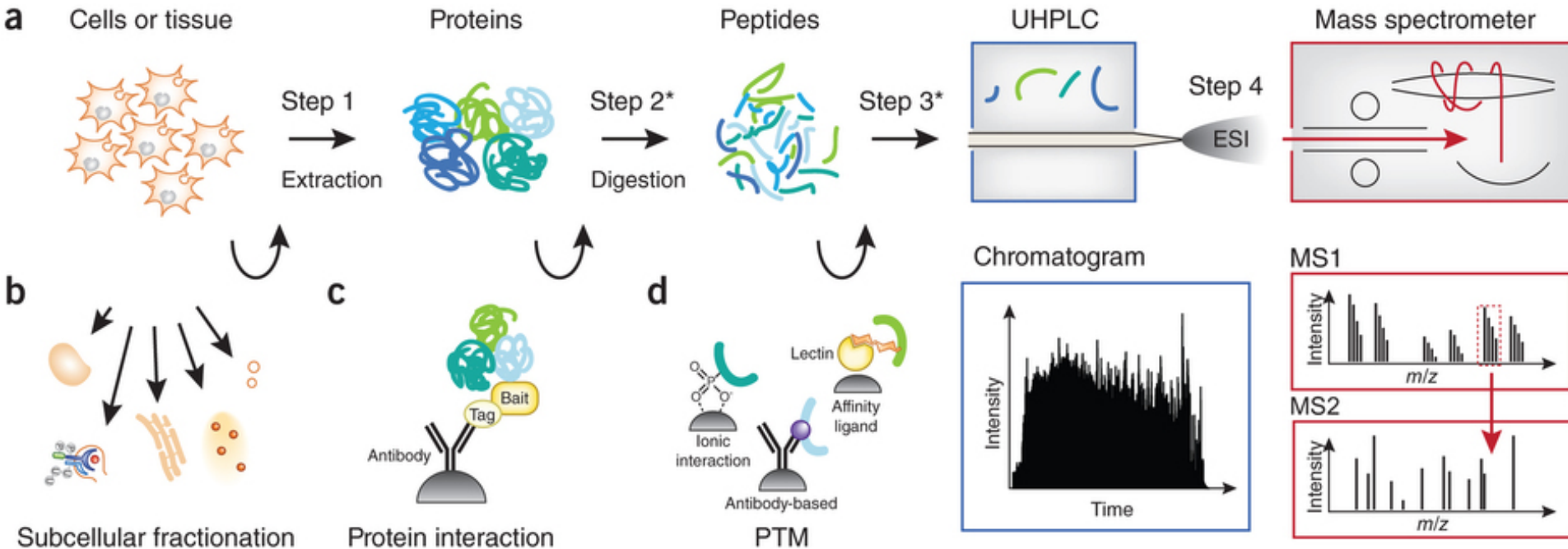


**Electrospray Tip - 20x**



**Electrospray Tip - 400x**

# Proteomics Workflow



## Considerations:

- Chemical nature of peptides of interest (PTM or un-modified)
- Complexity and dynamic range of mixture

# RADIO FREQUENCY TWO DIMENSIONAL QUADRUPOLE LINEAR ION TRAP

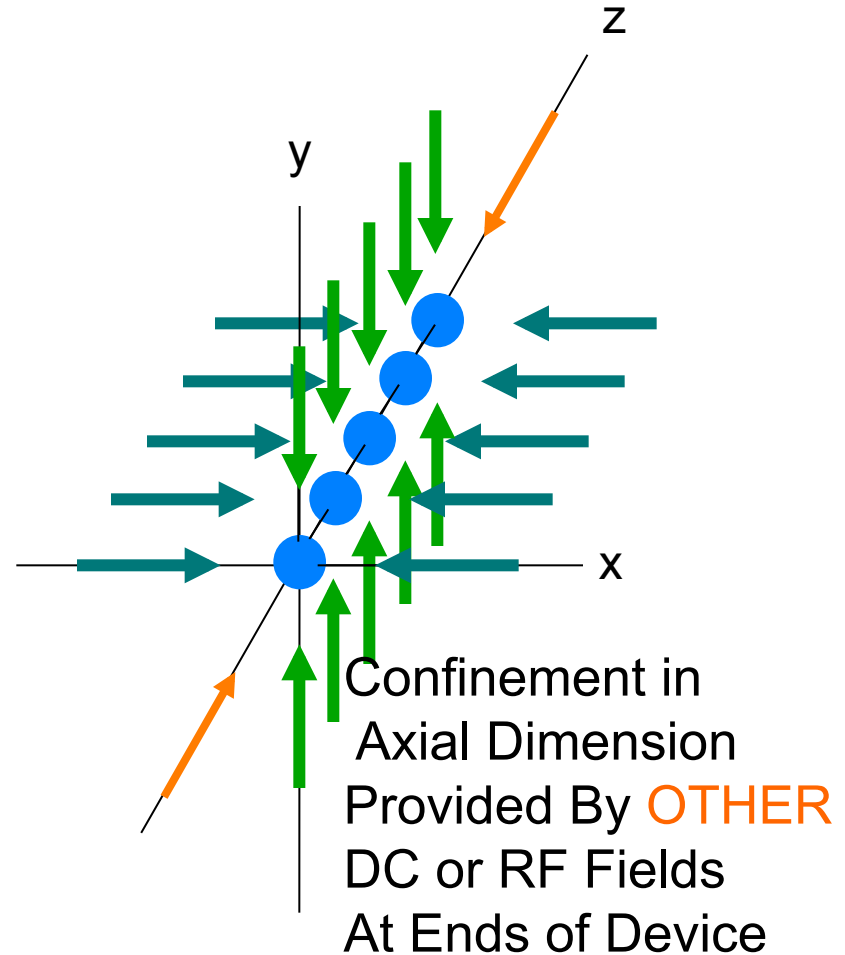
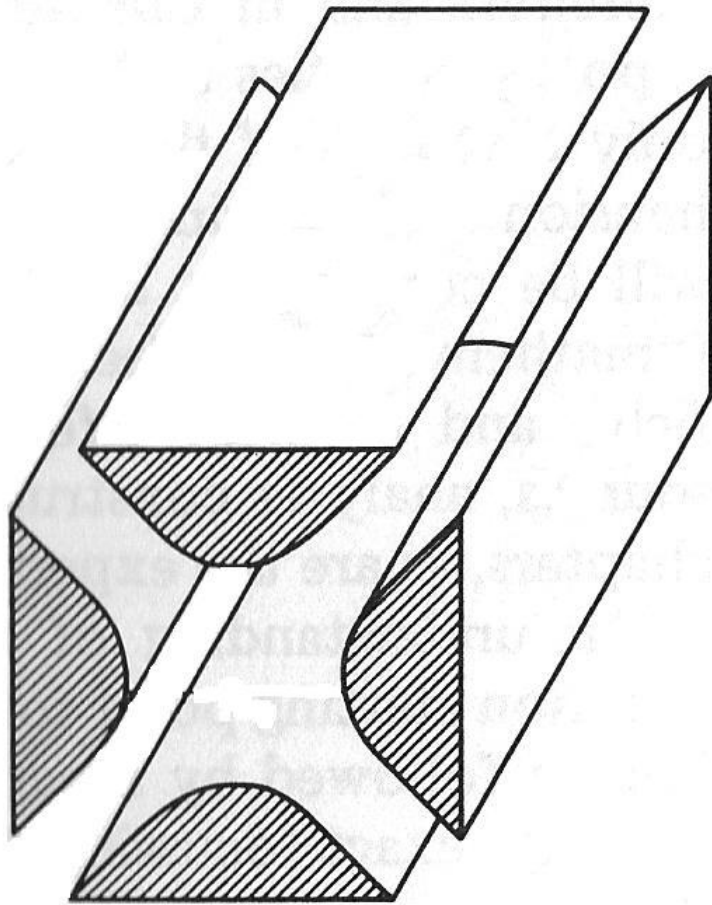
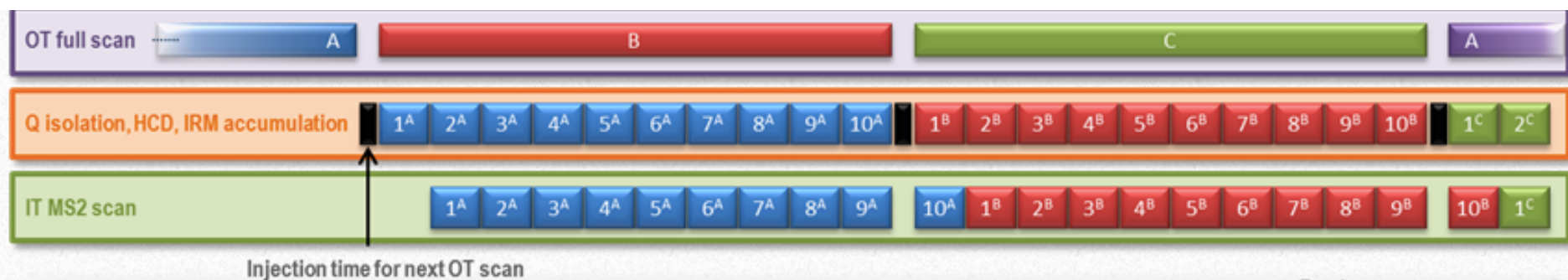
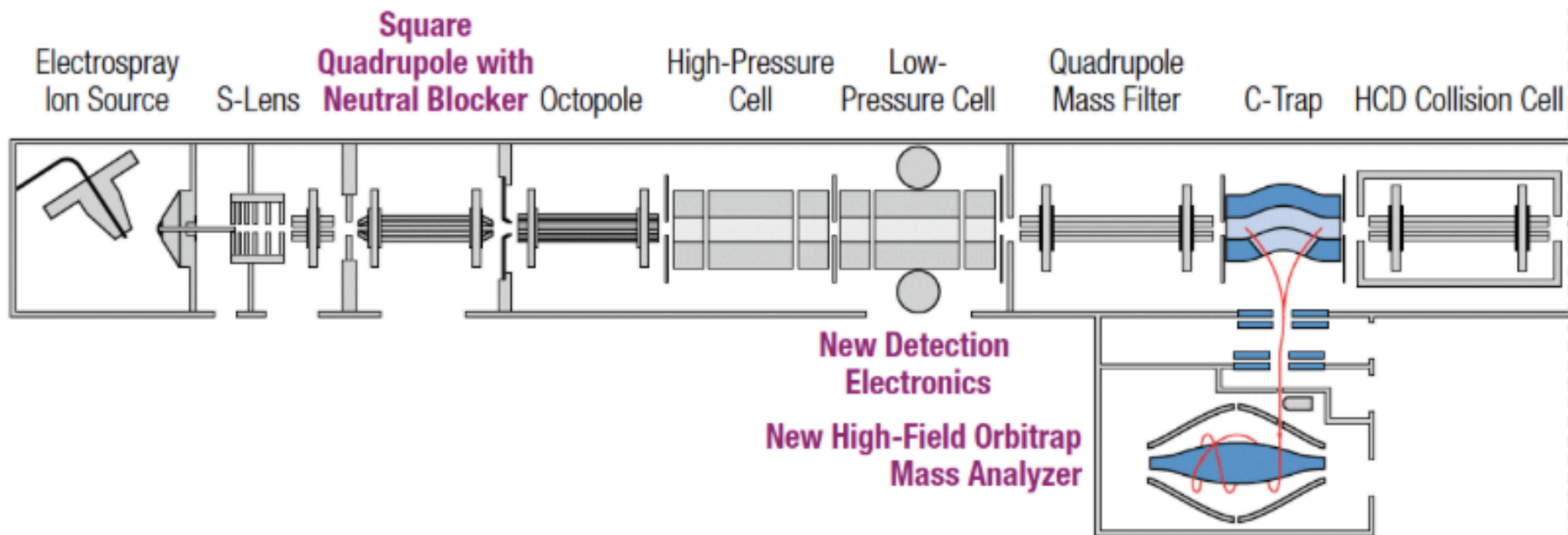


Figure From  
Quadrupole Mass Spectrometry and Its Applications  
P.H. Dawson Ed., Reprinted AIP Press 1995



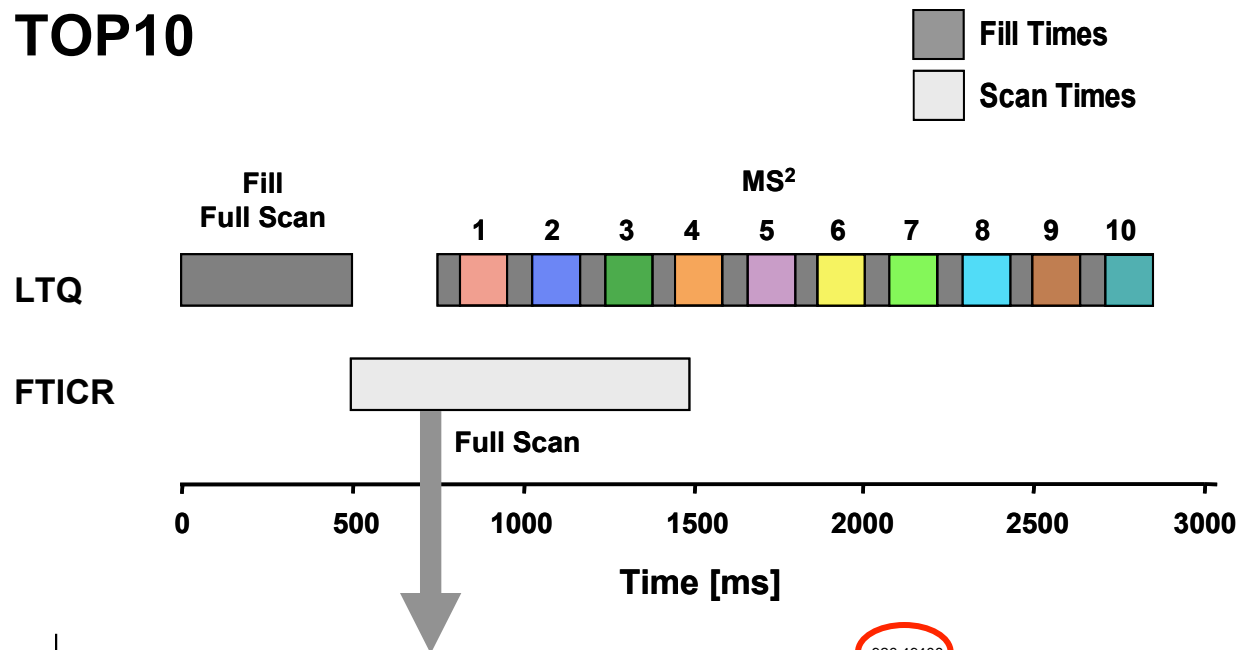
# Mass spec schematic and duty cycle



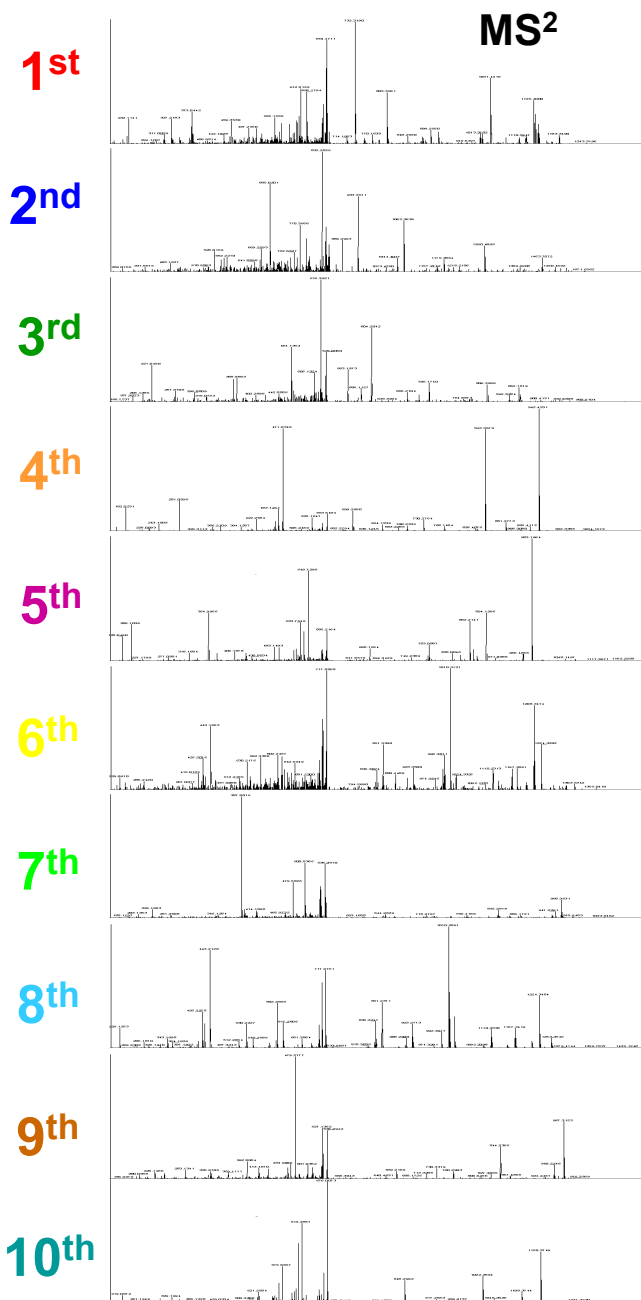
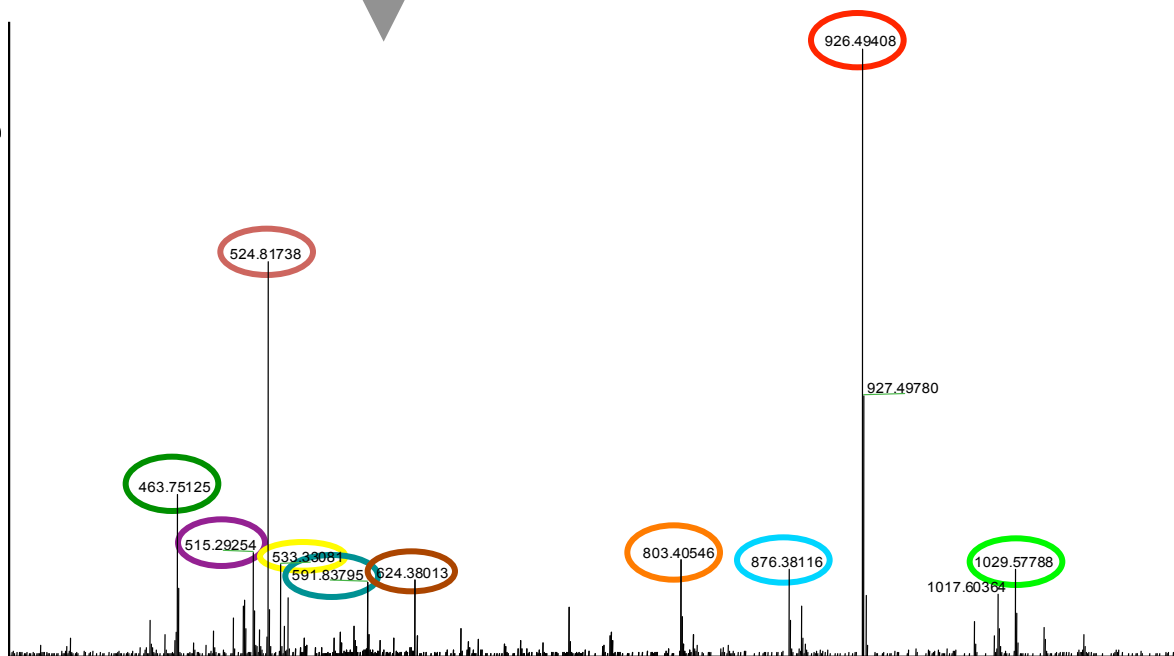


“shotgun sequencing”

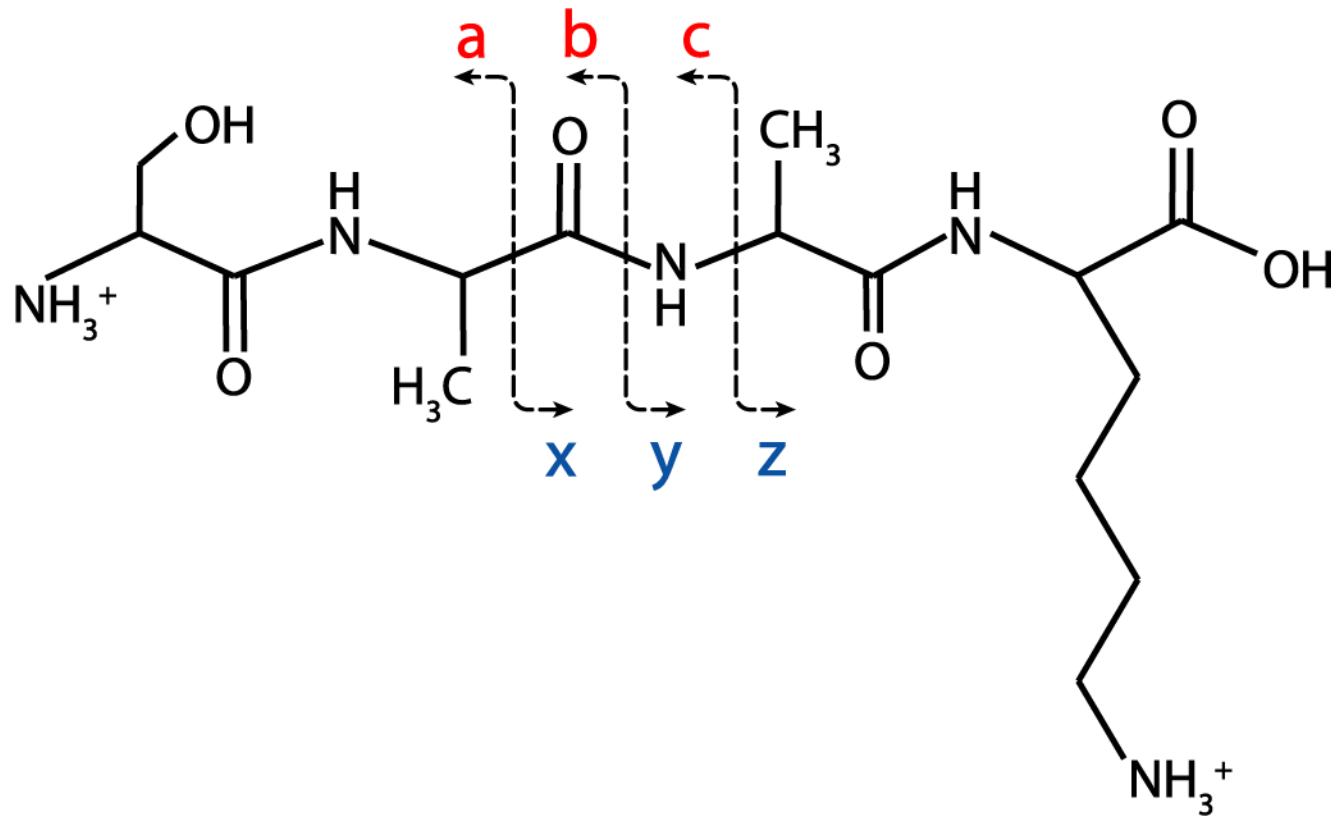
TOP10

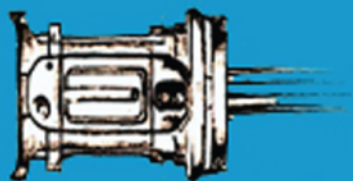


Relative Intensity



# Fragmentation nomenclature

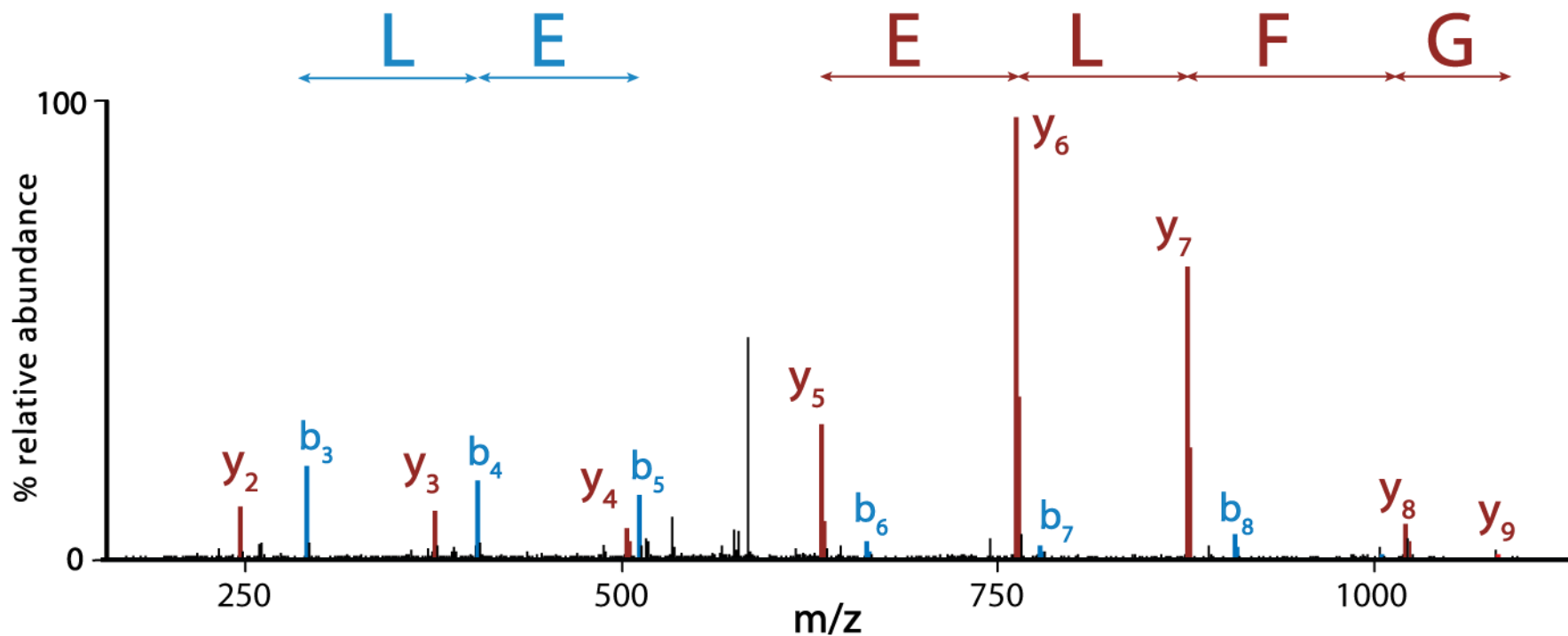




# Peptide Sequencing (MS/MS)

collision-activated dissociation (CAD)

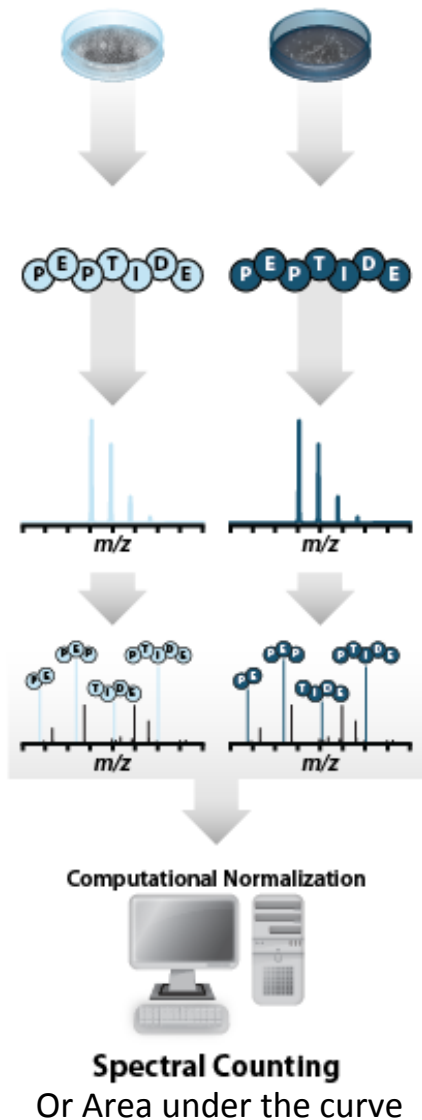
b <sup>+</sup>	<u>88</u>	<u>145</u>	<u>292</u>	<u>405</u>	<u>534</u>	<u>663</u>	<u>778</u>	<u>907</u>	<u>1020</u>	<u>1166</u>
	<b>S</b>	<b>G</b>	<b>F</b>	<b>L</b>	<b>E</b>	<b>E</b>	<b>D</b>	<b>E</b>	<b>L</b>	<b>K</b>
y <sup>+</sup>	<u>1166</u>	<u>1080</u>	<u>1022</u>	<u>875</u>	<u>762</u>	<u>633</u>	<u>504</u>	<u>389</u>	<u>260</u>	<u>147</u>



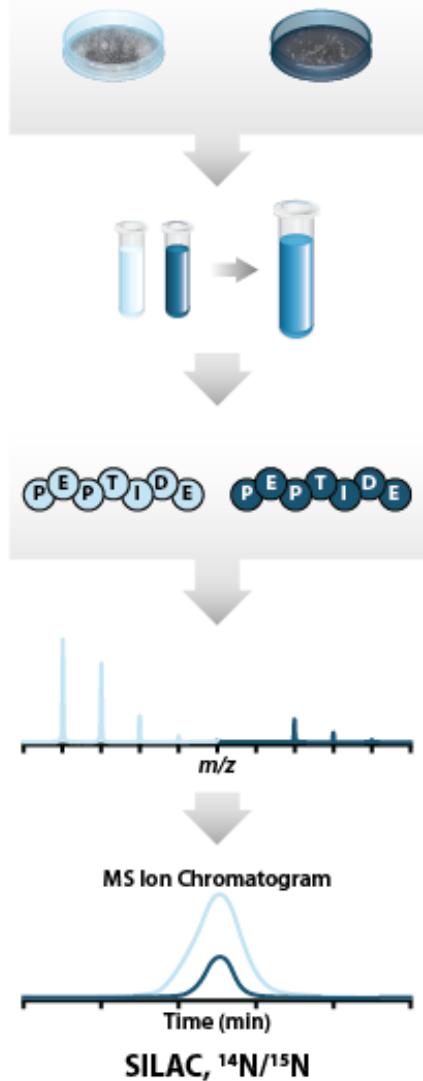
# Mass spec operation animation:

# Label-Based Quantitation

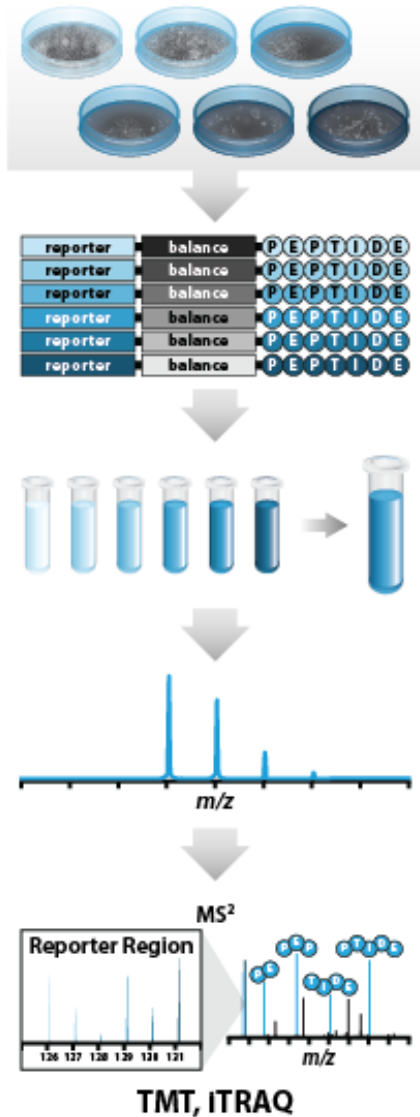
## A) Label Free



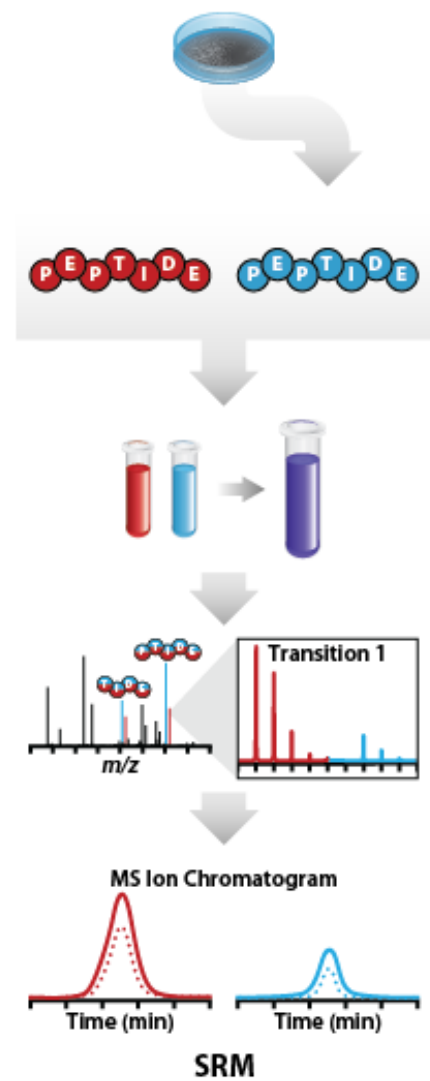
## B) Metabolic labeling



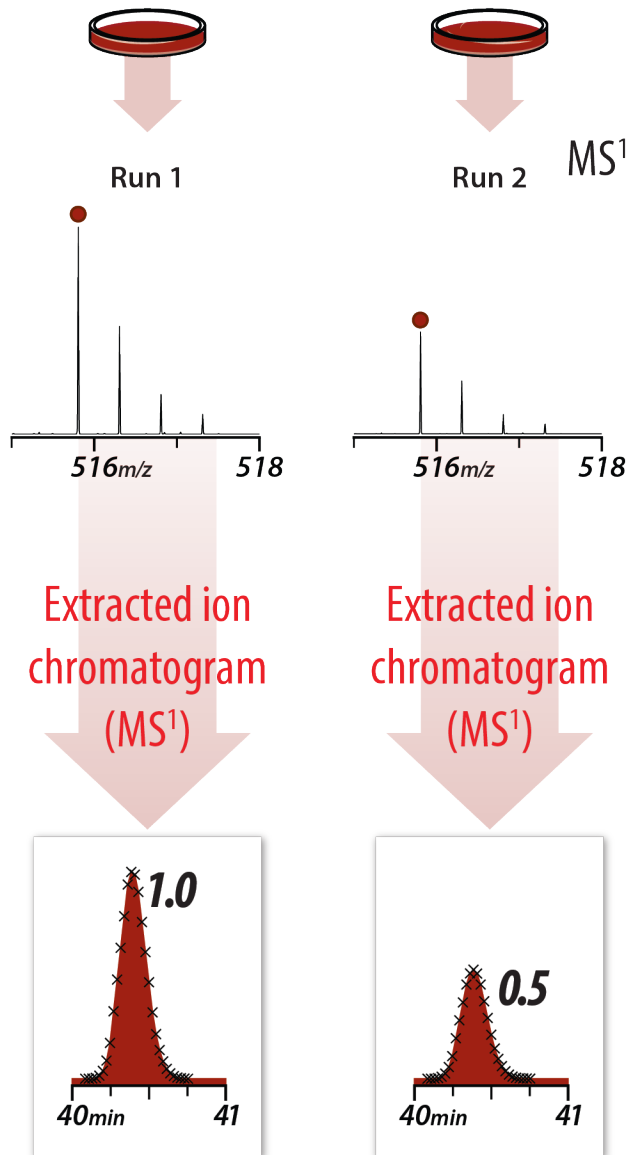
## C) Isobaric tagging



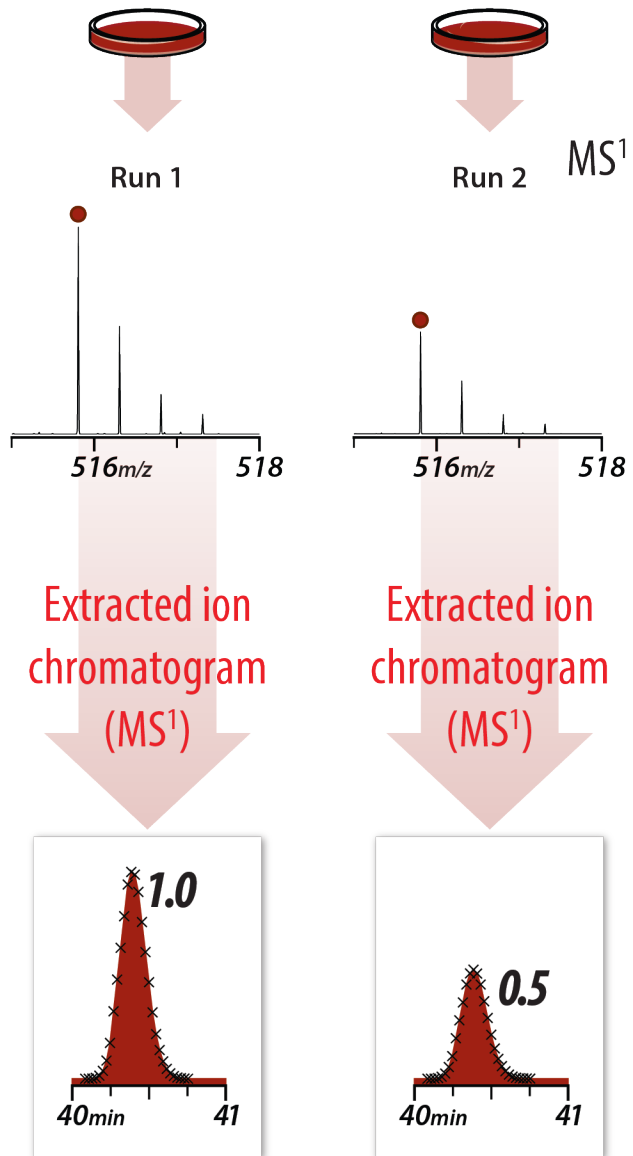
## D) SRM



# Label free quantitation AUC

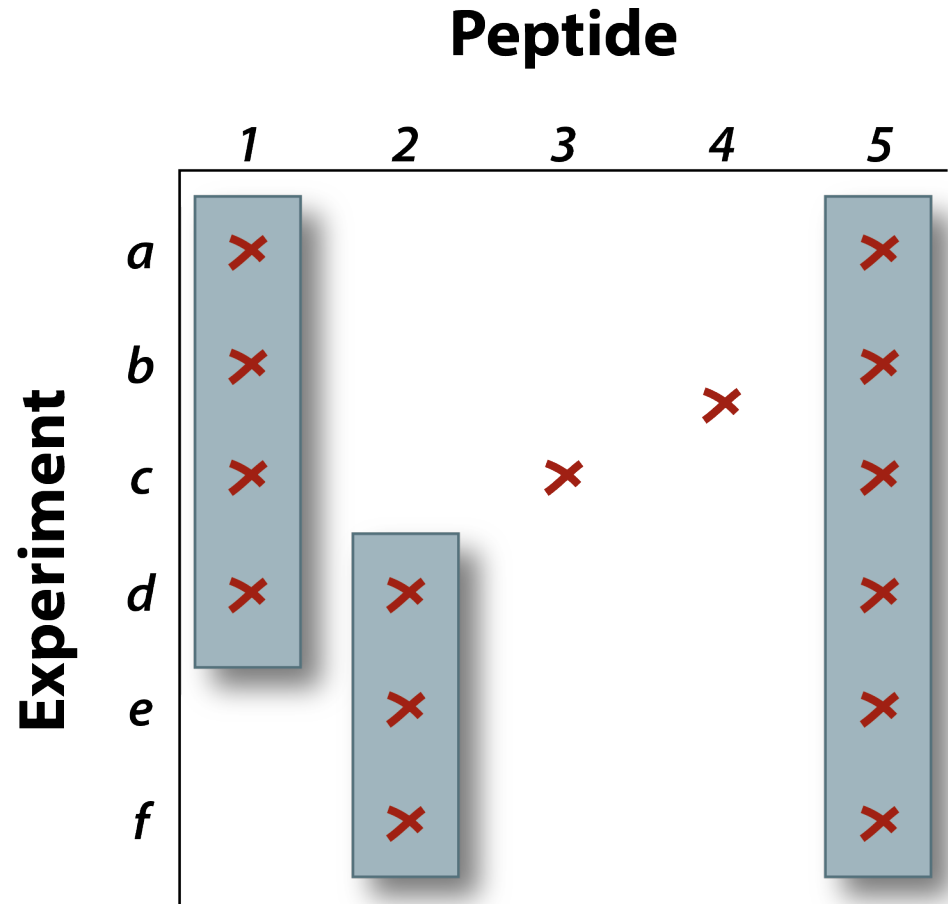
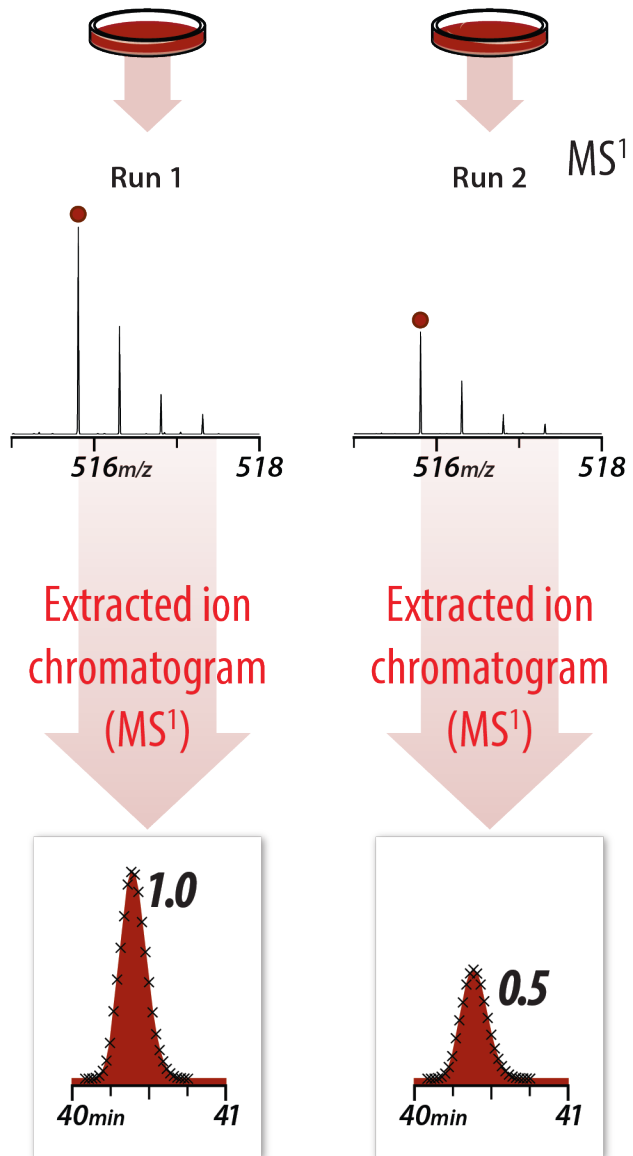


# Label free quantitation AUC



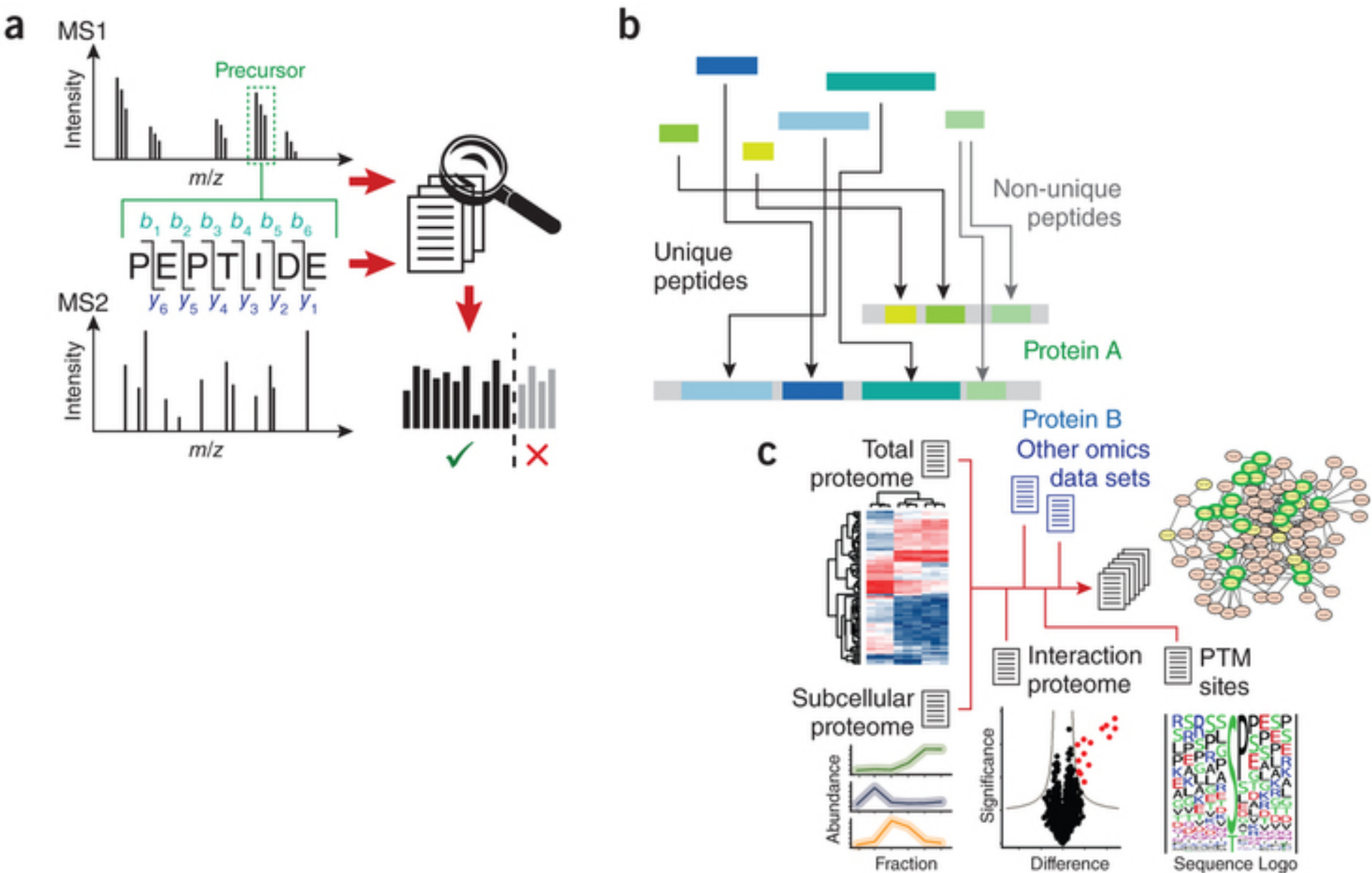
	Peptide				
	1	2	3	4	5
Experiment	a	×			×
	b	×		×	×
	c	×	×		×
	d	×			×
	e	×			×
	f	×			×

# Label free quantitation AUC

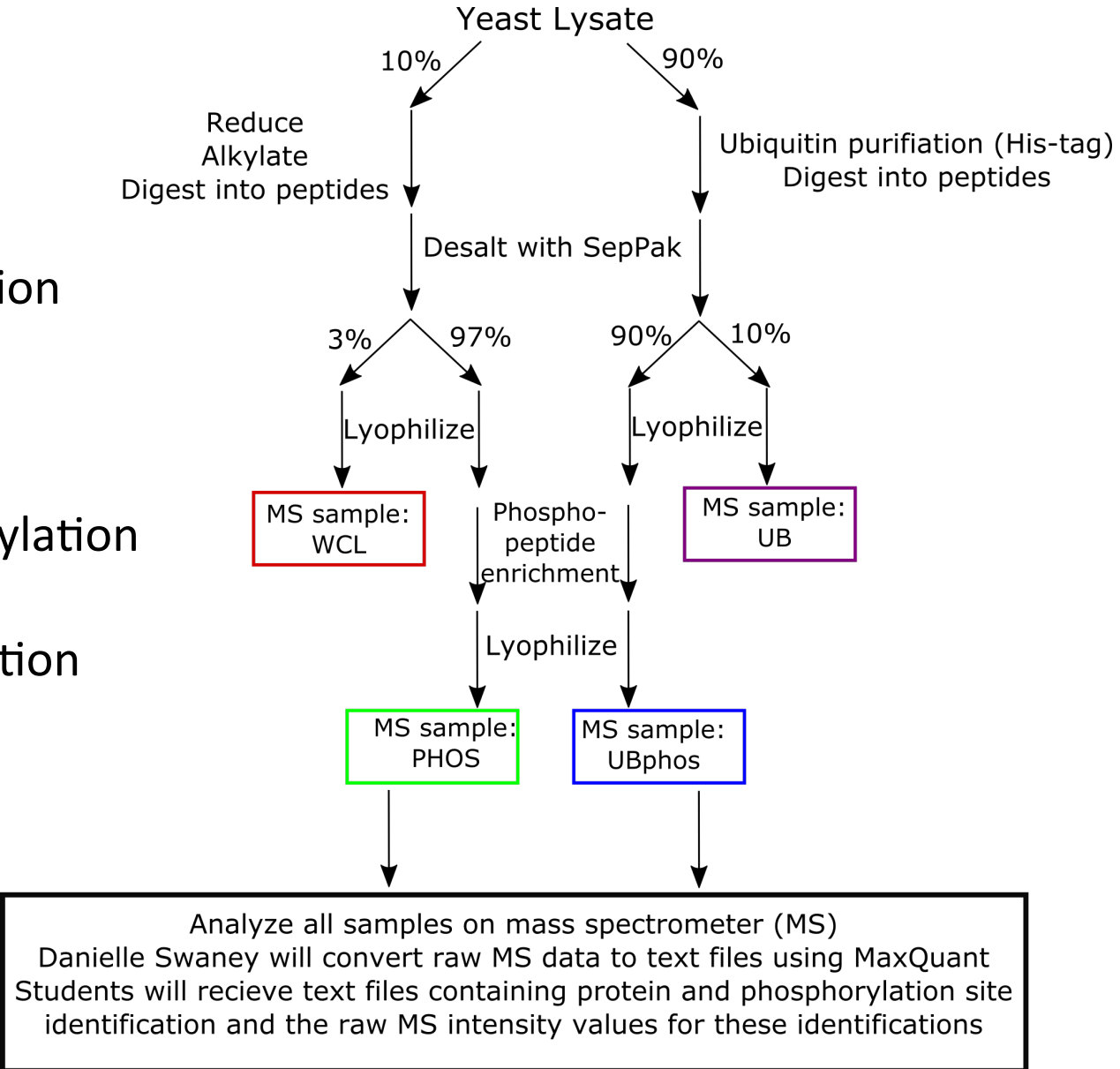




# What proteins did we identify? MaxQuant software



- Conditions:
  - Control
  - Kinase KO
  - Chemical perturbation
- Measurements:
  - Ubiquitin
  - Ubiquitin phosphorylation
  - Global proteome
  - Global phosphorylation



## \*\*\*PRO TIPS\*\*\*

- You are purifying a **MINORITY** population from a complex mixture.
- Focus on **REMOVING** as much of what you don't want from the sample as possible
- Worry less about maintaining 100% of your analyte of interest.
- Understand where your sample is.

